

(19)



Europäisches Patentamt

European Patent Office

Office européen des brevets



(11)

EP 0 754 756 A1

(12)

EUROPEAN PATENT APPLICATION

published in accordance with Art. 158(3) EPC

(43) Date of publication:
22.01.1997 Bulletin 1997/04

(21) Application number: 95909969.8

(22) Date of filing: 23.02.1995

(51) Int. Cl.⁶: C12N 15/53, C12P 13/08

(86) International application number:
PCT/JP95/00268

(87) International publication number:
WO 95/23864 (08.09.1995 Gazette 1995/38)

(84) Designated Contracting States:
DE DK ES FR GB IT NL

(30) Priority: 04.03.1994 JP 35019/94

(71) Applicant: Ajinomoto Co., Inc.
Tokyo 104 (JP)

(72) Inventors:
• SUGIMOTO, Masakazu
Ajinomoto Co., Inc.
Technology
ki-ku Kawasaki-shi Kanagawa 210 (JP)
• USUDA, Yoshihiro
Ajinomoto Co., Inc.
CentralResea
Kawasaki-shi Kanagawa 210 (JP)

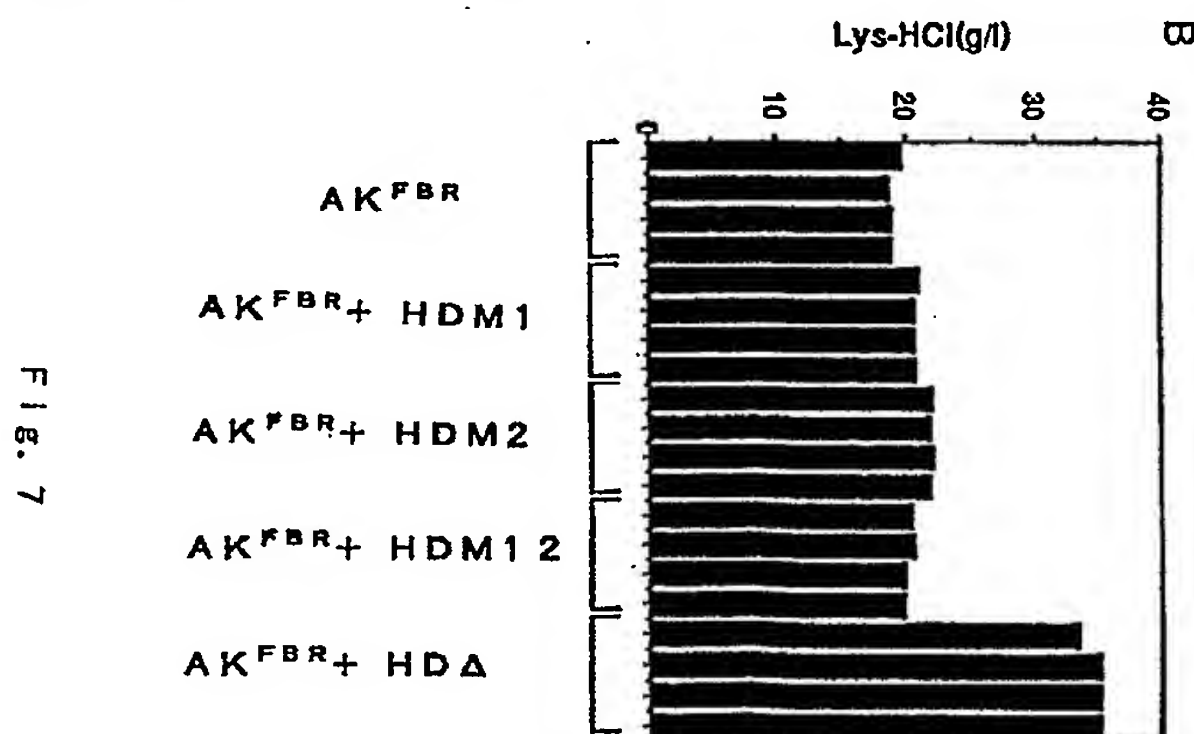
• SUZUKI, Tomoko
Ajinomoto Co., Inc.
CentralResearch
Kawasaki-shi Kanagawa 210 (JP)
• TANAKA, Akiko
Ajinomoto Co., Inc.
Central
Kawasaki-shi Kanagawa 210 (JP)
• MATSUI, Hiroshi
Ajinomoto Co., Inc.
Central
Kawasaki-shi Kanagawa 210 (JP)

(74) Representative: Hansen, Bernd, Dr. Dipl.-Chem.
et al
Hoffmann, Eitle & Partner,
Patentanwälte,
Arabellastrasse 4
81925 München (DE)

(54) PROCESS FOR PRODUCING L-LYSINE

(57) A coryneform bacterium having a high L-lysine productivity is provided by integrating a gene coding for a coryneform-origin aspartokinase released of feed-back inhibition caused by L-lysine and L-threonine into a DNA of a chromosome of a coryneform bacterium car-

rying attenuated homoserine dehydrogenase or a coryneform bacterium deficient in a homoserine dehydrogenase gene.



EP 0 754 756 A1

Description

Background of the Invention

The present invention relates to microbial industry, and in particular relates to a method of producing L-lysine by fermentation, and coryneform bacteria preferable for use in this production method.

L-lysine has been hitherto produced by fermentation using L-lysine-producing bacteria belonging to the genus Brevibacterium, Corynebacterium, Bacillus or Escherichia, which is synthesized in a biosynthesis system of any of these microorganisms from oxaloacetate through aspartate, aspartate β -aldehyde and so on. Various enzymes such as phosphoenol pyruvate carboxylase, aspartokinase and dihydrodipicolinate synthase participate in such an L-lysine biosynthesis pathway, however, many of these enzymes undergo feedback inhibition by L-lysine as a final product or by aspartic acid as an intermediate product. Thus when L-lysine is produced by fermentation, in order to improve the productivity, many mutant strains which do not undergo such inhibition are used.

For example, it is known that aspartokinase (hereinafter referred to as "AK") undergoes concerted inhibition by L-lysine and L-threonine synthesized in a branched pathway from the L-lysine synthesis pathway in coryneform bacteria belonging to the genera such as Brevibacterium and Corynebacterium. A mutant strain harboring AK which does not undergo the inhibition is used for L-lysine production (J. Gen. Appl. Microbiol., 16, 373-391 (1970)).

A mutant strain, which lacks homoserine dehydrogenase (hereinafter referred to as "HD") considered to be an enzyme having the greatest influence on L-lysine productivity, is also used for production of L-lysine by fermentation. This is attributed to the fact that L-threonine is not synthesized due to deficiency in HD to catalyze a reaction for producing L-homoserine from aspartate β -semialdehyde as a first reaction in a synthesis pathway inherent to L-threonine branching from the L-lysine synthesis pathway through aspartate β -semialdehyde, resulting in progress of the L-lysine synthesis reaction without inhibition of the AK activity. As such an HD deficient strain, an HD completely deficient strain of Corynebacterium glutamicum is known (Nakayama, K. et al.; J. Gen. Appl. Microbiol. 7(3), 145-154 (1961)).

In addition to the HD completely deficient strain as described above, a mutant strain harboring so-called leaky type HD is considered to be effective for L-lysine production, as well. The HD completely deficient strain cannot synthesize L-threonine and L-methionine, and thus it cannot grow unless these amino acids are present in a medium. On the contrary, if an HD leaky type strain can be obtained that harbors a leaky type HD which does not substantially exhibit activity so much to suppress L-lysine production but has HD activity a little, it becomes possible to make growth without addition of L-threonine and L-methionine to a medium, and it becomes convenient to prepare the medium.

Additionally, the leaky type HD has small affinity to aspartate β -semialdehyde as its substrate. Therefore, the HD leaky type strain synthesizes a considerable amount of aspartate β -semialdehyde for synthesizing L-threonine, L-methionine and L-isoleucine required for the growth. Aspartate β -semialdehyde synthesized in a considerable amount is consequently converted into L-lysine.

On the other hand, the HD completely deficient strain is considered to be still useful in that it completely suppresses production of L-threonine in amount, however, the deficiency in HD as a result of mutation has a possibility to restore the activity due to reverse mutation. Thus an HD deficient strain, in which such a possibility is extremely low with a destroyed HD gene, is considered to be more useful. A nucleotide sequence of an HD gene has been reported by Peoples et al. for Corynebacterium glutamicum (Peoples, O. P. et al., Molecular Microbiology, 2(1), 63-72 (1988)).

Since the HD leaky type strain and the HD deficient strain do not produce L-threonine, AK does not undergo feedback inhibition. Accordingly, it is expected that if the AK gene is amplified in cells of the HD leaky type strain and the HD deficient strain, the L-lysine synthesis reaction proceeds, and the L-lysine productivity is improved. It is further expected that L-lysine productivity is more improved by introducing, into coryneform bacteria, mutation of AK to avoid feedback inhibition by L-lysine and L-threonine in combination with leakage or deficiency of HD.

Disclosure of the Invention

The present invention has been made taking the aforementioned viewpoints into consideration, a task of which is to obtain an HD leaky type strain and an HD gene-destroyed strain, and provide an HD leaky type strain and an HD deficient strain with an amplified AK gene, and an HD leaky type strain and an HD gene-destroyed strain harboring AK which does not undergo feedback inhibition by L-lysine and L-threonine, in order to improve the L-lysine productivity of coryneform bacteria.

In order to solve the aforementioned task, the present inventors have obtained an HD leaky type mutant strain of Brevibacterium lactofermentum, isolated a wild type HD gene and a leaky type HD gene to clarify their structures, introduced the leaky type HD gene and a partially deleted HD gene into a wild strain of Brevibacterium lactofermentum, and thus created an L-lysine-producing strain having improved L-lysine productivity. The present inventors have succeeded in further improvement in L-lysine productivity by amplifying an AK gene in cells of the L-lysine-producing strain thus obtained, or by introducing a gene coding for AK which does not undergo feedback inhibition by L-lysine and L-threonine, and arrived at the present invention.

Namely, the present invention provides a DNA fragment which codes for homoserine dehydrogenase originating from a coryneform bacterium in which at least one of a 23rd leucine residue and a 104th valine residue as counted from its N-terminus is changed to another amino acid residue; a coryneform bacterium which harbors a gene coding for mutant homoserine dehydrogenase originating from a coryneform bacterium in which at least one of a 23rd leucine residue and a 104th valine residue as counted from its N-terminus is changed to another amino acid residue; and a coryneform bacterium which is transformed by integrating the aforementioned gene coding for mutant type homoserine dehydrogenase into chromosomal DNA by way of homologous recombination with a homoserine dehydrogenase gene on a chromosome of the coryneform bacterium.

In another aspect, the present invention provides a coryneform bacterium wherein its homoserine dehydrogenase gene is destroyed by integrating a DNA fragment coding for a part of homoserine dehydrogenase originating from a coryneform bacterium into chromosomal DNA by way of homologous recombination with a homoserine dehydrogenase gene on a chromosome of the coryneform bacterium. In still another aspect, the present invention provides a coryneform bacterium which harbors in its cells recombinant DNA constructed by ligating an aspartokinase gene originating from a coryneform bacterium with a vector autonomously replicable in cells of coryneform bacteria, and expresses no wild type homoserine dehydrogenase; and a coryneform bacterium which is transformed by integrating, into chromosomal DNA of the coryneform bacterium, a gene coding for aspartokinase originating from a coryneform bacterium with desensitized feedback inhibition by L-lysine and L-threonine, and expresses no wild type homoserine dehydrogenase. In still another aspect, the present invention provides a method of producing L-lysine comprising the steps of cultivating the coryneform bacterium described above in an appropriate medium, producing and accumulating L-lysine in a culture thereof, and collecting L-lysine from the culture.

In this specification, occasionally, a strain which produces wild type HD or wild type AK is referred to as "wild strain", HD having the leaky type mutation which scarcely exhibits substantial HD activity but has HD activity a little is merely referred to as "mutant type HD", AK having mutation not to undergo feedback inhibition by L-lysine and L-threonine is referred to as "mutant type AK", and a partially deleted HD gene is referred to as "deletion type HD gene". Further, the integration of recombinant DNA comprising a foreign HD gene or a foreign AK gene and a vector into chromosomal DNA by way of homologous recombination with an HD gene or an AK gene on host chromosomal DNA is referred to as "gene integration", and the achievement of a state in which the HD gene or the AK gene on the chromosome is replaced by the foreign HD gene or the foreign AK gene by allowing one copy of an HD gene or an AK gene to fall off together with the vector from a state in which the recombinant DNA is integrated into the chromosomal DNA is referred to as "gene replacement". Furthermore, a mutant strain harboring a mutant type HD gene or a strain subjected to gene replacement with a mutant type HD gene is merely referred to as "HD mutant strain", and a strain subjected to gene replacement with a partially deleted HD gene is referred to as "HD deficient strain", as well.

The coryneform bacteria referred to in the present invention is a group of microorganisms defined on page 599 in "Bergey's Manual of Determinative Bacteriology", eighth edition (1974), which reside in aerobic Gram-positive non-acid-fast rods having no spore-forming ability, including bacteria belonging to the genus Corynebacterium, bacteria belonging to the genus Brevibacterium having been hitherto classified into the genus Brevibacterium but united as bacteria belonging to the genus Corynebacterium at present, and bacteria belonging to the genus Brevibacterium closely relative to bacteria belonging to the genus Corynebacterium.

The HD mutant strain obtained according to the present invention is excellent in L-lysine productivity, and it can grow even when L-methionine and L-threonine, or L-homoserine is absent in a medium. The HD deficient strain of the present invention is excellent in L-lysine productivity because of no expression of the HD gene, and it can stably maintain this property.

Further, the HD mutant strain and the HD deficient strain with amplified AK gene, as well as the HD mutant strain and the HD deficient strain harboring the mutant type AK gene are more excellent in L-lysine productivity.

Detailed Description of the Invention

The present invention will be explained in detail below.

(1) Preparation of leaky type HD mutant strain and mutant type HD gene

A mutant strain which produces HD having leaky type mutation is obtained by performing a mutation treatment of a coryneform bacterium which produces wild type HD. For the mutation treatment of the coryneform bacterium, a treatment is conducted by using ultraviolet light irradiation or a mutating agent usually used for artificial mutation such as N-methyl-N'-nitro-N-nitrosoguanidine (NTG).

Bacterial cells after the mutation treatment were subjected to single colony isolation to select those producing the leaky type HD from each of colonies. Leaky type HD mutant strains can grow on a minimum medium, cannot grow on a minimum medium added with excessive L-methionine and L-threonine, but can grow on a minimum medium added with L-homoserine, or L-methionine and L-threonine. Thus they can be selected using the foregoing as a criterion

(Shiio, I. & Sano, K., J. G. A. M., 15, 267-287 (1969)). In order to confirm the fact that mutant strains thus obtained produce the leaky type HD, it is preferable to extract a crude enzyme solution from bacterial cells and compare the HD specific activity with that of wild type HD.

The enzyme activity of HD can be measured in accordance with, for example, a method of Kalinowski et al. (Kalinowski, J. et al., Mol. Gen. Genet., 224, 317-324 (1990)) using a crude enzyme solution prepared from bacterial cells in accordance with a method of Follettie et al. (Follettie, M. T. et al., Molecular Microbiology, 2, 53-62 (1988)).

In order to isolate the mutant type HD gene from the obtained leaky type HD mutant strains, chromosomal DNA is prepared from the leaky type HD mutant strains in accordance with, for example, a method of Saito and Miura (H. Saito and K. Miura, Biochem. Biophys. Acta, 72, 619 (1963)), and the HD gene is amplified by means of a polymerase chain reaction method (PCR; see White, T. J. et al., Trends Genet., 5, 185 (1989)). For DNA primers to be used for the amplification reaction, those complementary to both 3' terminals of a DNA double strand containing an entire or partial region of the HD gene are used. When only a partial region of the HD gene is amplified, it is necessary to screen a DNA fragment containing an entire region from a chromosomal DNA library using such DNA fragments as primers. When an entire region of the HD gene is amplified, a PCR reaction solution containing a DNA fragment including the amplified HD gene is subjected to agarose gel electrophoresis, followed by extraction of the objective DNA fragment. Thus the DNA fragment containing the HD gene can be recovered.

DNA primers may be appropriately prepared, for example, on the basis of a sequence known for Corynebacterium glutamicum (Peoples, O. P. et al, Molecular Microbiology, 2(1), 63-72 (1988)). Specifically, primers which can amplify a region comprising 1150 base pairs coding for the HD gene are preferable, and for example, two species of primers defined with SEQ ID NOS: 1 and 2 are suitable. The primer DNA can be synthesized in accordance with an ordinary method such as a phosphoamidite method (see Tetrahedron Letters, 22, 1859 (1981)) by using a commercially available DNA synthesizer (for example, DNA Synthesizer Model 380B produced by Applied Biosystems). Further, the PCR can be performed by using a commercially available PCR apparatus (for example, DNA Thermal Cycler Model PJ2000 produced by Takara Shuzo Co., Ltd.), using Taq DNA polymerase (supplied by Takara Shuzo Co., Ltd.) in accordance with a method designated by the supplier.

It is preferable for the mutant type HD gene amplified by the PCR method to be connected to vector DNA autonomously replicable in cells of Escherichia coli (hereinafter referred to as "E. coli", as well) and/or coryneform bacteria to prepare recombinant DNA which is introduced into E. coli cells, in order to facilitate following operations. The vector autonomously replicable in cells of E. coli is preferably a plasmid vector, preferably as those autonomously replicable in cells of a host, including, for example, pUC19, pUC18, pBR322, pHSG299, pHSG399, pHSG398, and RSF1010.

It is preferable that such a vector is inserted with a DNA fragment having an ability to make the plasmid autonomously replicable in coryneform bacteria which can be prepared, for example, from plasmids pAM330 (see Japanese Patent Laid-open No. 58-67699), pHM1519 (see Japanese Patent Laid-open No. 58-77895), pCG1 (see Japanese Patent Laid-open No. 57-134500), pCG2 (see Japanese Patent Laid-open No. 58-35197), pCG4 (see Japanese Patent Laid-open No. 57-183799), and pCG11 (see Japanese Patent Laid-open No. 57-183799). Thus the vector can be used as so-called shuttle vector which is autonomously replicable in both of E. coli and coryneform bacteria.

Such a shuttle vector is exemplified by the followings. Microorganisms harboring each of the vectors and deposition numbers of international deposition institutes are indicated in parentheses.

- pAJ655: Escherichia coli AJ11882 (FERM BP-136)
Corynebacterium glutamicum SR8201 (ATCC 39135)
- pAJ1844: Escherichia coli AJ11883 (FERM BP-137)
Corynebacterium glutamicum SR8202 (ATCC 39136)
- pAJ611: Escherichia coli AJ11884 (FERM BP-138)
- pAJ3148: Corynebacterium glutamicum SR8203 (ATCC 39137)
- pAJ440: Bacillus subtilis AJ11901 (FERM BP-140)

These vectors are obtained from deposited microorganisms as follows. Cells collected at the logarithmic growth phase are lysed with lysozyme and SDS to give a lysate from which a supernatant solution is obtained by centrifugation at 30,000 x g. Polyethylene glycol is added to the supernatant solution to perform fractional purification by means of cesium chloride-ethidium bromide equilibrium density gradient centrifugation.

In order to introduce a plasmid into E. coli for transformation, it is possible to use, for example, a method of D. M. Morrison (Methods in Enzymology, 68, 326 (1979)), or a method for treating recipient cells with calcium chloride to increase permeability of DNA (Mandel, M. and Higa, A., J. Mol. Biol., 53, 159 (1970)).

When the mutant type HD gene is isolated from the leaky type HD mutant strain, it is also obtained by preparing a chromosomal DNA library from the leaky type HD mutant strain using a plasmid vector or the like, selecting a strain harboring the mutant type HD gene from the library, and recovering recombinant DNA with the inserted mutant type HD gene from the selected strain. An example of a method for preparing the chromosomal library and selecting the strain harboring the mutant type HD gene from the library will be described below.

At first, a leaky type HD mutant strain is cultivated to obtain a culture. Any medium in which coryneform bacteria can grow is available for use. When L-threonine and L-methionine are contained in the medium in small amounts, it is preferable to add L-threonine and L-methionine, or L-homoserine beforehand. Next, bacterial cells are obtained by centrifuging the culture. Chromosomal DNA is obtained from the bacterial cells in accordance with, for example, a method of Saito and Miura (Biochem. Biophys. Acta, 72, 619 (1963)) or a method of K. S. Kirby (Biochem. J., 64, 405 (1956)).

In order to isolate the mutant type HD gene from the chromosomal DNA thus obtained, a chromosomal DNA library is prepared. At first, the chromosomal DNA is partially decomposed with an appropriate restriction enzyme to obtain a mixture of various fragments. A wide variety of restriction enzymes can be used by controlling the degree of cutting by controlling a period of time of the cutting reaction or the like. For example, Sau3AI is allowed to act on the chromosomal DNA to digest it at a temperature of not less than 30°C, preferably at 37°C at an enzyme concentration of 1-10 units/ml for various periods of time (1 minute to 2 hours).

Subsequently, the cut chromosomal DNA fragments are ligated with a vector autonomously replicable in E. coli cells to prepare recombinant DNA. Specifically, a restriction enzyme, which generates the same terminal nucleotide sequence as that by the restriction enzyme Sau3AI used for cutting the chromosomal DNA, for example, BamHI is allowed to act on the vector DNA to completely digest it under conditions of a temperature of not less than 30°C and an enzyme concentration of 1-100 units/ml for not less than 1 hour, preferably for 1-3 hours, to achieve cutting and cleavage. Next, the chromosomal DNA fragment mixture obtained as described above is mixed with the cleaved and cut vector DNA, on which DNA ligase, preferably T4 DNA ligase is allowed to act under conditions of a temperature of 4-16°C and an enzyme concentration of 1-100 units/ml for not less than 1 hour, preferably for 6-24 hours, to obtain recombinant DNA.

Using the obtained recombinant DNA, for example, E. coli K-12 strain is transformed to prepare a chromosomal DNA library. The transformation may be performed by, for example, a method of D. M. Morrison (Methods in Enzymology, 68, 326 (1979)), or a method for treating recipient cells with calcium chloride to increase permeability of DNA (Mandel, M. and Higa, A., J. Mol. Biol., 53, 159 (1970)).

In order to select a transformant strain harboring the mutant type HD gene from the obtained chromosomal DNA library, for example, an oligonucleotide probe may be synthesized on the basis of a sequence known for Corynebacterium glutamicum (Peoples, O. P. et al., Molecular Microbiology, 2(1), 63-72 (1988)), to perform colony hybridization using it. It is known that two kinds of E. coli HD genes (HD-1, HD-2) are present (Zakin, M. M. et al., J. B. C., 258, 3028-3031 (1983)), however, any of them has no region corresponding to about 100 amino acid residues on the C-terminal side of Corynebacterium glutamicum HD. Thus when a sequence to be used for the probe is selected from this region, it does not hybridize to the HD gene on E. coli chromosome, which is preferable. Recombinant DNA containing the mutant type HD gene can be isolated from transformed strains thus selected in accordance with, for example, a method of P. Guerry et al. (J. Bacteriol., 116, 1064 (1973)) or a method of D. B. Clewell (J. Bacteriol., 110, 667 (1972)).

Alternatively, a strain which produces the leaky type HD may be created by using a wild type HD gene cloned from a coryneform bacterium in the same manner as described above. At first, DNA containing a wild type HD gene or an HD gene having another mutation is subjected to an in vitro mutation treatment, and DNA after the mutation treatment is ligated with vector DNA adapted to a host to obtain recombinant DNA. The recombinant DNA is introduced into a host microorganism to obtain transformants, and one which expresses the leaky type HD is selected from the transformants. Alternatively, it is also available that DNA containing a wild type HD gene or an HD gene having another mutation is ligated with vector DNA adapted to a host to obtain recombinant DNA, thereafter the recombinant DNA is subjected to an in vitro mutation treatment, the recombinant DNA after the mutation treatment is introduced into a host microorganism to obtain transformants, and one which expresses the leaky type HD is selected from the transformants.

The agent for performing the in vitro mutation treatment of DNA is exemplified by hydroxylamine. Hydroxylamine is a treatment agent for chemical mutation which causes mutation from cytosine to thymine by changing cytosine to N⁴-hydroxycytosine.

The mutant type HD gene for use in the present invention is not especially limited provided that it codes for the leaky type HD, for which there are exemplified genes coding for HD having any of mutations in an amino acid sequence of wild type HD, including:

- (1) mutation to change a 23rd leucine residue from the N-terminal to an amino acid residue other than the leucine residue;
- (2) mutation to change a 104th valine residue from the N-terminal to an amino acid residue other than the valine residue; and
- (3) mutation to change the 23rd leucine residue from the N-terminal to an amino acid residue other than the leucine residue, and the 104th valine residue from the N-terminal to an amino acid residue other than the valine residue.

The amino acid sequence of wild type HD is herein specifically exemplified by an amino acid sequence of HD originating from a wild type strain of Brevibacterium lactofermentum shown in SEQ ID NO: 3 and SEQ ID NO: 4 in Sequence Listing.

With respect to the mutations described in the foregoing (1) to (3), mutation to change to a phenylalanine residue is exemplified for the 23rd leucine residue, and mutation to change to an isoleucine residue is exemplified for the 104th valine residue.

Any codon corresponding to the replaced amino acid residue is available especially regardless of its type provided that it codes for the amino acid residue. The amino acid sequence of harbored wild type HD may slightly differ depending on difference in bacterial species and bacterial strains. HD having such replacement, deletion or insertion of amino acid residues at positions irrelevant to the activity of the enzyme can be also used for the present invention.

For example, as will be described in Examples below, as a result of comparison of an amino acid sequence of HD originating from Brevibacterium lactofermentum 2256 strain (ATCC 13869) with an amino acid sequence reported for HD of Corynebacterium glutamicum (Peoples, O. P. et al., Molecular Microbiology, 2(1), 63-72 (1988)), it has been clarified that a 148th amino acid residue from the N-terminal is a glycine residue in HD of Corynebacterium glutamicum, while it is an alanine residue in HD of Brevibacterium lactofermentum. It is expected that the leaky type HD is obtained by introducing any of the aforementioned mutations (1) to (3) even in the case of HD of Corynebacterium glutamicum as described above.

(2) Preparation of wild type AK gene and mutant type AK gene

The wild type AK gene for use in the present invention can be prepared from wild strains of coryneform bacteria. A gene, which codes for AK in which cumulative feedback inhibition by L-lysine and L-threonine is substantially desensitized, can be prepared from a mutant strain in which cumulative feedback inhibition to the AK activity by L-lysine and L-threonine is substantially desensitized. Such a mutant strain can be obtained from a group of cells having been subjected to a mutation treatment applied to, for example, a wild strain of coryneform bacteria by using an ordinary mutation treatment method such as ultraviolet light irradiation or a treatment with a mutating agent such as N-methyl-N'-nitro-N-nitrosoguanidine (NTG). For measuring the AK activity, it is possible to use a method described by Miyajima, R. et al. in The Journal of Biochemistry (1968), 63(2), 139-148.

With respect to donor bacteria for the AK gene, a wild strain ATCC 13869 of Brevibacterium lactofermentum, and an L-lysine-producing bacterium AJ3463 (FERM P-1987) derived from the ATCC 13869 strain by a mutation treatment are most preferable donor bacteria.

In order to isolate the AK gene from the coryneform bacteria, chromosomal DNA is prepared in accordance with, for example, a method of Saito and Miura (H. Saito and K. Miura, Biochem. Biophys. Acta, 72, 619 (1963)), and the AK gene is amplified by means of a polymerase chain reaction method (PCR; see White, T. J. et al., Trends Genet., 5, 185 (1989)).

For DNA primers to be used for the amplification, those complementary to both 3' terminals of a DNA double strand containing an entire or partial region of the AK gene are used. When only a partial region of the AK gene is amplified, it is necessary to perform screening by amplifying a DNA fragment containing an entire region from a gene library using DNA fragments of the region as primers. When an entire region is amplified, the DNA fragment is subjected to agarose gel electrophoresis, followed by excision of an objective band. Thus the DNA fragment containing the AK gene can be recovered.

For DNA primers, single strand DNA's of 23 mer and 21 mer represented by 5'-TCGCGAAGTAGCACCTGTCACTT-3' (SEQ ID NO: 5 in Sequence Listing) and 5'-ACGGAATTCAATCTTACGGCC-3' (SEQ ID NO: 6 in Sequence Listing) are most suitable to amplify a region of about 1643 bp based on, for example, a sequence known for Corynebacterium glutamicum (see Molecular Microbiology (1991), 5(5), 1197-1204; Mol. Gen. Genet. (1990), 224, 317-324). The DNA can be synthesized in accordance with an ordinary method using a phosphoramidite method (see Tetrahedron Letters (1981), 22, 1859) by using a DNA synthesizer Model 380B produced by Applied Biosystems. The PCR can be performed by using DNA Thermal Cycler Model PJ2000 produced by Takara Shuzo Co., Ltd., using Taq DNA polymerase in accordance with a method designated by the supplier.

It is preferable for the mutant type AK gene amplified by the PCR method to be connected to vector DNA autonomously replicable in cells of E. coli and/or coryneform bacteria to prepare recombinant DNA which is introduced into E. coli cells, in order to facilitate following operations. The vector autonomously replicable in cells of E. coli is preferably a plasmid vector, preferably as those autonomously replicable in cells of a host, including, for example, pUC19, pUC18, pBR322, pHSG299, pHSG399, pHSG398, and RSF1010.

When these vectors are inserted with a DNA fragment having an ability to make the plasmid autonomously replicable in coryneform bacteria, they can be used as so-called shuttle vectors which are autonomously replicable in both of E. coli and coryneform bacteria. In order to introduce a plasmid into E. coli for transformation, it is possible to use, for example, a method of D. M. Morrison (Methods in Enzymology, 68, 326 (1979)), or a method for treating recipient cells with calcium chloride to increase permeability of DNA (Mandel, M. and Higa, A., J. Mol. Biol., 53, 159 (1970)).

The wild type AK gene is obtained by isolating the AK gene from the AK wild strain as described above, and the mutant type AK gene is obtained by isolating the AK gene from the AK mutant strain.

The mutant type AK gene to be used for the present invention is not especially limited provided that it codes for AK

in which cumulative feedback inhibition by L-lysine and L-threonine is desensitized. However, its mutation may be exemplified, with respect to the amino acid sequence of the wild type AK, such that a 279th alanine residue from the N-terminal is changed to an amino acid residue other than alanine and other than acidic amino acids in α -subunit, and a 30th alanine residue is changed to an amino acid residue other than alanine and other than acidic amino acids in β -subunit.

5 The amino acid sequence of the wild type AK is herein specifically exemplified by an amino acid sequence defined in SEQ ID NO: 10 in Sequence Listing for the α -subunit, and an amino acid sequence defined in SEQ ID NO: 12 in Sequence Listing for the β -subunit.

The aforementioned amino acid residue other than alanine and other than acidic amino acids is exemplified by threonine, arginine, cysteine, phenylalanine, proline, serine, tyrosine and valine residues.

10 Any codon corresponding to the amino acid residue to be replaced is available especially regardless of its type provided that it codes for the amino acid residue. It is postulated that the amino acid sequence of harbored wild type AK may slightly differ depending on difference in bacterial species and bacterial strains. AK having such replacement, deletion or insertion of amino acid residues at positions irrelevant to the activity of the enzyme can be also used for the present invention.

15 (3) Preparation of HD mutant strain and HD deficient strain

The HD mutant strain is obtained as described in (1) by performing a treatment of a coryneform bacterium which produces wild type HD with ultraviolet light irradiation or a mutating agent, and selecting a strain which produces mutant type HD from bacterial cells subjected to the mutation treatment. The HD mutant strain which expresses no wild type HD is also obtained by introducing a mutant type HD gene isolated from the HD mutant strain thus obtained into cells of a wild type coryneform bacterium, and performing gene replacement by way of homologous recombination with an HD gene on chromosome.

25 The mutant type HD gene may be replaced with the HD gene on the host chromosome as follows (see Fig. 1). Namely, a temperature-sensitive replication origin originating from *Brevibacterium lactofermentum*, the mutant type HD gene, and a marker gene for exhibiting resistance to a drug such as chloramphenicol are inserted into a plasmid vector to prepare recombinant DNA. The recombinant DNA is used to transform a coryneform bacterium, transformed strains are cultivated at a temperature at which the temperature-sensitive replication origin does not operate, and then they are cultivated in a medium containing the drug. Thus a transformed strain, in which the recombinant DNA is integrated into chromosomal DNA, is obtained.

30 The strain with the recombinant DNA integrated into the chromosome causes recombination with an HD gene sequence originally existing on the chromosome, in which two fused genes of the chromosomal HD gene and the mutant type HD gene are inserted into the chromosome in a state of interposing other portions of the recombinant DNA (vector portion, temperature-sensitive replication origin, and drug resistance marker). Therefore, the wild type HD is dominant in this state, and thus equivalent growth to that of the wild strain is exhibited in a minimum medium.

35 Next, in order to allow only the mutant type HD gene to remain on the chromosomal DNA, one copy of an HD gene is allowed to fall off together with the vector portion (including the temperature-sensitive replication origin and the drug resistance marker) by recombination of the two HD genes. For example, the strain with the integration on the chromosome is cultivated, and cultivated bacterial cells are spread and cultivated on a solid plate medium containing no drug. Grown colonies are replicated and cultivated on a solid plate medium containing the drug, and drug-sensitive strains are obtained. The fact that the vector portion falls off from chromosomes of the obtained drug-sensitive strains is confirmed by Southern hybridization, and the fact that the mutant type HD is expressed is confirmed.

45 When the gene replacement is performed by using an HD gene coding for a part of HD, that is a partially deleted HD gene, instead of the aforementioned mutant type HD gene, an HD deficient strain in which its chromosomal HD gene is replaced with the partially deleted HD gene is obtained.

As described in Example 1 below, it is postulated that a region on the N-terminal side in HD participates in the activity. Therefore, the site to be deleted in the HD gene is exemplified by a region on the N-terminal side, for example, a region within 350 amino acids, for example, a region of 100th to 200th amino acids or 250th to 350th amino acids from the N-terminal. Since the HD gene is located in the same operon as that of homoserine kinase existing downstream therefrom, it is preferable that the promoter site of the HD gene is not deleted so as not to inhibit expression of homoserine kinase.

55 Introduction of the recombinant DNA into cells of coryneform bacteria is possible by using a method in which recipient cells are treated with calcium chloride to increase permeability of DNA as reported for *E. coli* K-12 (Mandel, M. and Higa, A., *J. Mol. Biol.*, 53, 159 (1970)), or a method in which introduction is performed in a growth stage so that cells can incorporate DNA (so-called competent cells) as reported for *Bacillus subtilis* (Duncan, C. H., Wilson, G. A. and Young, F. E., *Gene*, 1, 153 (1977)). Alternatively, it is also possible to perform introduction into recipients for recombinant DNA after converting the DNA recipients into protoplasts or spheroplasts which easily incorporate recombinant DNA, as known for *Bacillus subtilis*, actinomycetes and yeasts (Chang, S. and Choen, S. N., *Molec. Gen. Genet.*, 168, 111 (1979); Bibb, M. J., Ward, J. M. and Hopwood, O. A., *Nature*, 274, 398 (1978); Hinnen, A., Hicks, J. B. and Fink, G.

R., Proc. Natl. Acad. Sci. USA, 75, 1929 (1978)).

In the protoplast method, a sufficiently high frequency can be obtained even by the method used for Bacillus subtilis described above. It is of course possible to utilize a method in which DNA is incorporated into protoplasts of the genus Corynebacterium or Brevibacterium in the presence of polyethylene glycol or polyvinyl alcohol and divalent metal ion as described in Japanese Patent Laid-open No. 57-183799. An equivalent result is obtained even by a method in which incorporation of DNA is facilitated by addition of carboxymethyl cellulose, dextran, Ficoll, Pluronic F68 (Serva Co.) instead of polyethylene glycol or polyvinyl alcohol.

Further, recombinant DNA can be introduced into recipients belonging to bacteria of the genus Brevibacterium or Corynebacterium by using an electric pulse method (Sugimoto et al., Japanese Patent Laid-open No. 2-207791).

The wild type coryneform bacteria into which the mutant type HD gene or the deletion type HD gene is introduced are exemplified by bacteria belonging to the genus Corynebacterium, bacteria belonging to the genus Brevibacterium having been hitherto classified into the genus Brevibacterium but united as bacteria belonging to the genus Corynebacterium at present, and bacteria belonging to the genus Brevibacterium closely relative to bacteria belonging to the genus Corynebacterium. Especially, glutamate-producing bacteria belonging to the genus Corynebacterium (Brevibacterium) are most preferable in the present invention. Examples of wild strains of the glutamate-producing bacteria belonging to the genus Corynebacterium (Brevibacterium) includes the followings. These wild strains as well as strains added with a property of L-lysine production to such strains can be used for the present invention in the same manner.

<u>Corynebacterium acetoacidophilum</u>	ATCC 13870
<u>Corynebacterium acetoglutamicum</u>	ATCC 15806
<u>Corynebacterium callunae</u>	ATCC 15991
<u>Corynebacterium glutamicum</u>	ATCC 13032
	ATCC 13060
<u>Brevibacterium divaricatum</u>	ATCC 14020
<u>Brevibacterium lactofermentum</u>	ATCC 13869
<u>Corynebacterium lilium</u>	ATCC 15990
<u>Corynebacterium melassecola</u>	ATCC 17965
<u>Brevibacterium saccharolyticum</u>	ATCC 14066
<u>Brevibacterium immariophilum</u>	ATCC 14068
<u>Brevibacterium roseum</u>	ATCC 13825
<u>Brevibacterium flavum</u>	ATCC 13826
<u>Brevibacterium thiogenitalis</u>	ATCC 19240
<u>Microbacterium ammoniaphilum</u>	ATCC 15354

The coryneform bacteria which can be used for the present invention includes mutant strains having glutamate productivity or those lacking glutamate productivity, in addition to the wild strains having glutamate productivity as described above. At present, various artificial mutant strains of coryneform glutamate-producing bacteria are used as L-lysine-producing bacteria, and these strains can be also used for the present invention. Such artificial mutant strains include the followings: AEC (S-(2-aminoethyl)-cysteine) resistant mutant strains; mutant strains which require amino acid such as L-homoserine (Japanese Patent Publication Nos. 48-28078 and 56-6499); mutant strains which exhibit resistance to AEC and require amino acids such as L-leucine, L-homoserine, L-proline, L-serine, L-arginine, L-alanine, and L-valine (United States Patent Nos. 3708395 and 3825472); L-lysine-producing mutant strains which exhibit resistance to DL- α -amino- ϵ -caprolactam, α -amino-lauryllactam, aspartate-analog, sulfa drug, quinoid, N-lauroylleucine; L-lysine-producing mutant strains which exhibit resistance to inhibitors of oxaloacetate decarboxylase or respiratory system enzymes (Japanese Patent Laid-open Nos. 50-53588, 50-31093, 52-102498, 53-9394, 53-86089, 55-9783, 55-9759, 56-32995 and 56-39778, and Japanese Patent Publication Nos. 53-43591 and 53-1833); L-lysine-producing mutant strains which require inositol or acetic acid (Japanese Patent Laid-open Nos. 55-9784 and 56-8692); L-lysine-producing mutant strains which exhibit sensitivity to fluoropyruvic acid or temperature not less than 34°C (Japanese Patent Laid-open Nos. 55-9783 and 53-86090); and mutant strains of Brevibacterium or Corynebacterium which exhibit resistance to ethylene glycol and produce L-lysine (see United States Patent Application Serial No. 333455).

(4) Amplification of AK gene in HD mutant strain or HD deficient strain

AK undergoes feedback inhibition in the case of co-existence of L-lysine and L-threonine. However, coryneform bacteria which do not express wild type homoserine dehydrogenase cannot produce L-threonine, and thus AK does not undergo feedback inhibition.

Therefore, it is expected that the L-lysine productivity is improved if the AK gene is amplified in cells of coryneform bacteria which do not express wild type homoserine dehydrogenase. It is further expected that the L-lysine productivity is more improved if an inhibition-desensitized type AK gene is used as the AK gene to be amplified because feedback inhibition is more reduced.

The coryneform bacteria for introduction of the AK gene which do not express homoserine dehydrogenase are exemplified by the HD mutant strain or the HD deficient strain obtained as described in (3) above. However, the effect of improvement in L-lysine productivity is obtained owing to the amplification of the AK gene in the same manner even when an HD completely deficient strain obtained by a mutation treatment is used.

In order to amplify the AK gene or the mutant type AK gene in cells of such coryneform bacteria which do not express wild type homoserine dehydrogenase, the coryneform bacteria may be transformed with recombinant DNA comprising the AK gene or the mutant type AK gene and a vector autonomously replicable in cells of coryneform bacteria.

The vector used herein may be any one provided that it is autonomously replicable in cells of coryneform bacteria. Specifically, there may be exemplified pAJ655, pAJ1844, pAJ611, pAJ3148 and pAJ440 described above.

The method for transforming coryneform bacteria is exemplified by a method in which recipient cells are treated with calcium chloride to increase permeability of DNA as reported for *E. coli* K-12 (Mandel, M. and Higa, A., *J. Mol. Biol.*, **53**, 159 (1970)), or a method in which introduction is performed in a growth stage so that cells can incorporate DNA (so-called competent cells) as reported for *Bacillus subtilis* (Duncan, C. H., Wilson, G. A. and Young, F. E., *Gene*, **1**, 153, (1977)). Alternatively, it is also possible to perform introduction into recipients for recombinant DNA after converting the DNA recipients into protoplasts or spheroplasts which easily incorporate recombinant DNA, as known for *Bacillus subtilis*, actinomycetes and yeasts (Chang, S. and Choen, S. N., *Molec. Gen. Genet.*, **168**, 111 (1979); Bibb, M. J., Ward, J. M. and Hopwood, O. A., *Nature*, **274**, 398 (1978); Hinnen, A., Hicks, J. B. and Fink, G. R., *Proc. Natl. Acad. Sci. USA*, **75**, 1929 (1978)).

Additionally, the stability of recombinant DNA in a host can be improved by allowing the vector to harbor a marker gene such as drug resistance, or a gene to supplement auxotrophy of the host.

A promoter inherent to the AK gene may be used exactly as it is for expressing the AK gene or the mutant type AK gene. However, it is also available that a promoter of another gene which operates in coryneform bacteria is used to ligate it with a DNA sequence coding for AK or mutant type AK.

(5) Introduction of mutant type AK gene into chromosomal DNA of HD mutant strain or HD deficient strain

The L-lysine productivity can be improved by performing amplification of the AK gene in cells of the HD mutant strain or the HD deficient strain as described in (4) above. However, in order to increase the stability of the AK gene introduced into the HD mutant strain or the HD deficient strain, it is preferable to integrate the AK gene into chromosomal DNA. It is preferable herein to use a mutant type AK gene as the AK gene to be integrated into chromosomal DNA.

In order to integrate the mutant type AK gene into chromosomal DNA of a host, integration of the gene may be performed in the same manner as the mutant type HD gene or the deletion type HD gene. Namely, a temperature-sensitive replication origin originating from *Brevibacterium lactofermentum*, the mutant type AK gene, and a marker gene for providing resistance to a drug such as chloramphenicol are inserted into a plasmid vector to prepare recombinant DNA. The recombinant DNA is used to transform a coryneform bacterium, transformed strains are cultivated at a temperature at which the temperature-sensitive replication origin does not operate, and then they are cultivated in a medium containing the drug. Thus a transformed strain, in which the recombinant DNA is integrated into chromosomal DNA, is obtained.

The strain with the recombinant DNA integrated into the chromosomal DNA causes recombination with an AK gene sequence originally existing on the chromosome, in which two fused genes of the chromosomal AK gene and the mutant type AK gene are inserted into the chromosome in a state of interposing other portions of the recombinant DNA (vector portion, temperature-sensitive replication origin, and drug resistance marker). The mutant type AK is dominant in this state, and thus the phenotype is the mutant type. Therefore, the strain integrated with the gene may be used as it is. However, when approximately the same sequences are aligned in parallel on the chromosomal DNA, recombination may take place again, and one of the AK genes is apt to fall off. Accordingly, it is preferable to obtain a gene-replaced strain in which only the mutant type AK gene remains on the chromosomal DNA. Namely, one copy of the AK gene is allowed to fall off together with the vector portion (including the temperature-sensitive replication origin and the drug resistance marker) by recombination of the two AK genes. For example, the strain with the integration on the chro-

mosome is cultivated, and cultivated bacterial cells are spread and cultivated on a solid plate medium containing no drug. Grown colonies are replicated and cultivated on a solid plate medium containing the drug, and drug-sensitive strains are obtained. The fact that the vector portion falls off from chromosomes of the obtained drug-sensitive strains is confirmed by Southern hybridization, and the fact that the mutant type AK is expressed is confirmed.

No problem arises even when the wild type AK gene remains on the chromosomal DNA in a complete form, as being different from the case of the gene replacement using the mutant type HD gene or the deletion type HD gene. Accordingly, the mutant type AK gene may be integrated at a site other than that for the AK gene on the chromosomal DNA.

(6) Production of L-lysine

L-lysine can be produced and accumulated in a culture by cultivating, in an appropriate medium, the HD mutant strain, the HD deficient strain, the strains of these types in which the AK gene is amplified, or the HD mutant strain or the HD deficient strain into which the mutant type AK gene is integrated.

The medium to be used includes an ordinary medium containing a carbon source, a nitrogen source, inorganic ions and optionally other organic components.

As the carbon source, it is possible to use sugars such as glucose, lactose, galactose, fructose, or starch hydrolysate; or organic acids such as fumaric acid, citric acid or succinic acid.

As the nitrogen source, it is possible to use inorganic ammonium salts such as ammonium sulfate, ammonium chloride or ammonium phosphate; organic nitrogen such as soybean hydrolysate; ammonia gas; or aqueous ammonia.

It is desirable to allow required substances such as vitamin B₁ and L-homoserine or yeast extract to be contained in appropriate amounts as organic trace nutrients. Other than the above, potassium phosphate, magnesium sulfate, iron ion, manganese ion and the like are added in small amounts, if necessary.

When the HD deficient strain is used, L-threonine and L-methionine, or L-homoserine is added to the medium in appropriate amount(s).

Cultivation is preferably carried out under an aerobic condition for 16-72 hours. The cultivation temperature is preferably controlled at 25°C to 37°C, and pH is preferably controlled at 5-7 during cultivation. Inorganic or organic, acidic or alkaline substances as well as ammonia gas or the like can be used for pH adjustment. Collection of L-lysine from a cultivated liquor may be carried out by combining an ordinary ion exchange resin method, a precipitation method and other known methods.

Brief Description of the Drawings

Fig. 1 is a conceptual view of gene integration and gene replacement;

Fig. 2 is a view of comparison of amino acid sequences of HD genes of various microorganisms;

Fig. 3 is a view of comparison of amino acid sequences of HD genes of various microorganisms (continued);

Fig. 4 shows process of construction of p399AK9B and p399AKYB;

Fig. 5 shows L-lysine productivity and OD after cultivation of HD mutant strain and HD deficient strain;

Fig. 6 shows L-lysine productivity and OD after cultivation of HD mutant strain and HD deficient strain in which AK gene is amplified; and

Fig. 7 shows L-lysine productivity and OD after cultivation of HD mutant strain and HD deficient strain in which mutant type AK gene is integrated into chromosome.

Description of Preferred Embodiments

The present invention will be more concretely explained below with reference to Examples.

Example 1: Analysis of Wild Type HD Gene, Leaky Type HD Gene, and Inhibition-Desensitized Type HD Gene

Leaky type HD mutant strains and a mutant strain for producing HD with desensitized feedback inhibition by L-threonine were created from a wild strain of Brevibacterium lactofermentum. A wild type HD gene, leaky type HD genes and a inhibition-desensitized type HD gene were isolated from the wild and mutant strains, and analysis of their structures was performed. Brevibacterium lactofermentum AJ12036 strain (FERM BP-734) was used as the wild strain. Brevibacterium lactofermentum AJ12472 and AJ12937 strains were used as the leaky type HD mutant strains. Brevibacterium lactofermentum AI6080 strain was used as the inhibition-desensitized type HD mutant strain. These mutant strains were obtained as follows.

The AJ12036 strain is a strain obtained by deleting an originally existing plasmid, pAM330 from Brevibacterium lactofermentum 2256 strain (ATCC 13869), which produces wild type HD with respect to HD.

The AJ12472 and AJ12937 strains are strains obtained from Brevibacterium lactofermentum 2256 strain (ATCC

13869) as a result of repeated breeding by mutation using L-lysine productivity as an index, which produce leaky type HD's. The AI6080 strain is a strain obtained from Brevibacterium lactofermentum 2256 strain (ATCC 13869) as a result of repeated breeding by mutation using L-threonine productivity as an index, which produces inhibition-desensitized type HD.

(1) Amplification of HD gene by PCR method

A nucleotide sequence of the HD gene has been reported for Corynebacterium glutamicum (Peoples, O. P. et al, Molecular Microbiology, 2(1), 63-72 (1988)). It was speculated that the similarity of each HD gene sequence might be high between Brevibacterium lactofermentum and Corynebacterium glutamicum. Thus synthetic primer DNA's were prepared on the basis of the sequence of Corynebacterium glutamicum for use in the PCR method.

Chromosomal DNA's were prepared in accordance with an ordinary method from Brevibacterium lactofermentum AJ12036, AJ12472, AJ12937 and AI6080 strains. In order to amplify DNA fragments of about 1500 bp containing HD genes from these chromosomal DNA's, a DNA synthesizer Model 1381A (ABI Ltd.) was used to synthesize two species of primers of a 5' side primer H1 ((841)5'-CTGGG AAGGTGAATCGAATTT-3'(860), SEQ ID NO: 1 in Sequence Listing) and a 3' side primer H2 ((2410)5'-TCCGAGGTTTGCA GAAGATC-3'(2391), SEQ ID NO: 2 in Sequence Listing). Numbers in the parentheses indicate positions in the nucleotide sequence published by People et al. (Peoples, O. P. et al, Molecular Microbiology, 2(1), 63-72 (1988)). Obtained synthetic primers were purified by reversed phase HPLC.

The PCR was performed with a composition shown below using a PCR amplification apparatus (DNA Thermal Cycler PJ2000 produced by Takara Shuzo Co., Ltd.) and a PCR kit (Takara GeneAmp™ kit produced by Takara Shuzo Co., Ltd.).

Table 1

Component	Concentration	Blended amount
Primer H1	0.25 μ M	25 pmol
Primer H2	0.25 μ M	25 pmol
dATP, dGTP, dTTP, dCTP	each 200 μ M	20 nmol
<u>Taq</u> DNA polymerase	2.5 U/100 μ L	0.5 μ L (5 U/ μ L)
Chromosomal DNA		1 μ g
10 x reaction buffer		10 μ L
Water		balance
		(total amount: 100 μ L)

Conditions for denaturation of DNA, annealing of DNA, and polymerase reaction in the PCR were at 94°C for 1 minute, at 37°C for 2 minutes and at 75°C for 3 minutes respectively, and transition between each of the temperatures was performed for 1 second. DNA was amplified by repeating the reaction cycle by 25 cycles. As a result of confirmation of sizes of amplification reaction products thus obtained by using agarose gel electrophoresis, amplification of DNA fragment of about 1.4 Kbp was observed.

Thus the DNA fragments amplified from the chromosomal DNA of each of the AJ12036, AJ12472, AJ12937 and AI6080 strains were respectively cut with a restriction enzyme KpnI to obtain DNA fragments which were inserted into a KpnI site of a vector plasmid pHSG399 (see Takeshita, S. et al., Gene (1987), 61, 63-74) to obtain recombinant DNA's. The recombinant DNA's containing amplified fragments originating from the AJ12036, AJ12472, AJ12937 and AI6080 strains were designated as pHDW, pHDMI, pHDMII and pHDMIII, respectively. Each of the plasmids was introduced into E. coli JM109 strain to obtain transformants.

(2) Determination of nucleotide sequences of HD genes and analysis of mutation points

(1) Comparison of nucleotide sequences of wild and mutant type HD genes

Nucleotide sequences of the HD gene fragments of Brevibacterium lactofermentum AJ12036, AJ12472, AJ12937 and AI6080 strains obtained as described above were determined by the dideoxy method.

A determined nucleotide sequence of the wild type HD gene of the AJ12036 strain, and an amino acid sequence

deduced from the sequence are shown in SEQ ID NO: 3 in Sequence Listing. Further, the amino acid sequence is shown in SEQ ID NO: 4 in Sequence Listing. As a result of comparison of the sequence with the sequence of the HD gene of *Corynebacterium glutamicum* reported by Peoples et al. (Peoples, O. P. et al, *Molecular Microbiology*, 2(1), 63-72 (1988)), nucleotides were different at 4 places, and one of them was different at the amino acid level. The different points are shown below using the sequence of the HD gene of *Corynebacterium glutamicum* as a standard.

- (1) $^{531}\text{G} \rightarrow \text{C}$ ($^{148}\text{Gly} \rightarrow ^{148}\text{Ala}$)
- (2) $^{1222}\text{G} \rightarrow \text{C}$
- (3) $^{1318}\text{G} \rightarrow \text{T}$
- (4) $^{1324}\text{C} \rightarrow \text{G}$

Such a diversity observed among HD genes of wild type strains of coryneform bacteria do not affect the HD activity, and the sequence of the HD gene of *Corynebacterium glutamicum* may be treated as equivalent of the sequence of the HD gene of *Brevibacterium lactofermentum* shown in SEQ ID NO: 3.

As a result of comparison of the nucleotide sequence of the wild type HD gene and the amino acid sequence deduced from the sequence of the AJ12036 strain with nucleotide sequences of the HD genes and amino acid sequences of the AJ12472, AJ12937 and AI6080 strains, mutation points were found for the AJ12472 strain at 2 places, for AJ12937 at 1 place, and for AI6080 at 1 place, all accompanying amino acid replacement. Further, it was found that exactly the same mutation was commonly present in the HD genes of the AJ12472 and AJ12937 strains at 1 place. Each of the mutation points is shown below.

Table 2

Bacterial strain	Difference in nucleotide sequence	Mutation in amino acid residue
AJ12472 strain	$^{155}\text{G} \rightarrow \text{T}$ $^{398}\text{G} \rightarrow \text{A}$	$^{23}\text{Leu} \rightarrow \text{Phe}$ $^{104}\text{Val} \rightarrow \text{Ile}$
AJ12937 strain	$^{398}\text{G} \rightarrow \text{A}$	$^{104}\text{Val} \rightarrow \text{Ile}$
AI6080 strain	$^{1266}\text{C} \rightarrow \text{T}$	$^{393}\text{Ser} \rightarrow \text{Phe}$

Hereinafter, the mutation point of $^{155}\text{G} \rightarrow \text{T}$ ($^{23}\text{Leu} \rightarrow \text{Phe}$) is referred to as "mutation point 1", the mutation point of $^{398}\text{G} \rightarrow \text{A}$ ($^{104}\text{Val} \rightarrow \text{Ile}$) is referred to as "mutation point 2", and the mutation point of $^{1266}\text{C} \rightarrow \text{T}$ ($^{393}\text{Ser} \rightarrow \text{Phe}$) is referred to as "mutation point 3".

(2) Comparison of HD amino acid sequences and mutation points of *Brevibacterium lactofermentum*, *Bacillus subtilis* and *E. coli*

It is known that two kinds of HD genes (HD-1, HD-2) are present in *E. coli*, and any of them constitutes bifunctional enzyme with AK (Zakin, M. M. et al., *J. B. C.*, 258, 3028-3031 (1983)). Further, a nucleotide sequence of an HD gene of *Bacillus subtilis* has been also determined (Parsot, C. and Cohen, G. N., *J. B. C.*, 263(29), 14654-14660 (1988)). Comparison of these amino acid sequences with the amino acid sequence of the wild type HD of *Brevibacterium lactofermentum* is shown in Figs. 2 and 3.

According to the result, it is understood that most of sites having high homology are located in a region on the N-terminal side, and that sites having high homology are concentrated in a region having amino acid numbers of 100-230 especially in the amino acid sequence of HD of *Brevibacterium lactofermentum*. It is postulated that the active region of HD exists on the N-terminal side according to the aforementioned fact, the fact that the two mutation points of HD of *Brevibacterium lactofermentum* are located within about 100 amino acid residues from the N-terminal, especially the mutation point 1 is located at a position of 23 amino acid residues from the N-terminal, and the two mutation points are amino acid residues having high conservation with respect to HD-1 and HD-2 of *E. coli*, HD of *Bacillus subtilis*, and HD of *Brevibacterium lactofermentum*, and the fact that no sequence corresponding to about 100 amino acid residues on the C-terminal side of HD of *Brevibacterium lactofermentum* is present in HD-1 and HD-2 of *E. coli*.

On the other hand, nucleotide sequences of HD genes of *Corynebacterium glutamicum* with desensitized inhibition by L-threonine have been published. Namely, Sahm et al. have reported replacement of one 68th amino acid from the C-terminal due to point mutation (Reinscheid, D. J. et al., *J. Bacteriol.*, 173(10), 3228-3230 (1991)), and Sinskey et al. have reported change in 17th amino acid and followings due to frame shift on account of point mutation, and deletion

of 7th amino acid and followings from the C-terminal (Archer, J. A. C. et al., Gene, 107, 53-59 (1991)). Further, the mutation of amino acid residue was at a position of 53th amino acid residue from the C-terminal in the inhibition-desensitized HD of the Brevibacterium lactofermentum A16080 strain. Furthermore, the region on the C-terminal side which does not exist in HD-1 and HD-2 of E. coli exists in HD of Bacillus subtilis which undergoes feedback inhibition by L-threonine in the same manner as HD of Brevibacterium lactofermentum. Accordingly, it is speculated that the region of HD relating to the feedback inhibition by L-threonine exists on the C-terminal side.

Example 2: Preparation and Analysis of Wild Type AK Gene and Mutant Type AK Gene

(1) Construction of wild type and mutant type AK genes and plasmids containing them

Chromosomal DNA's were prepared in accordance with an ordinary method from Brevibacterium lactofermentum 2256 strain (ATCC 13869) and an L-lysine-producing mutant strain AJ3463 (FERM P-1987) obtained from the 2256 strain by a mutation treatment. AK genes were amplified from the chromosomal DNA's in accordance with the PCR method (polymerase chain reaction; see White, T. J. et al., Trends Genet., 5, 185 (1989)). DNA primers used in the amplification were based on a sequence known for Corynebacterium glutamicum (see Molecular Microbiology (1991), 5(5), 1197-1204, and Mol. Gen. Genet. (1990), 224, 317-324). In order to amplify a region of about 1643 bp coding for the AK gene, single strand DNA's of 23 mer and 21 mer having sequences of 5'-TCGCGAAGTAGCACCTGTCACTT-3' (SEQ ID NO: 5) and 5'-ACGGAATTCAATCTTACGGCC-3' (SEQ ID NO: 6) were synthesized. The DNA's were synthesized by using a DNA synthesizer Model 380B produced by Applied Biosystems, Ltd. in accordance with an ordinary method using the phosphoamidite method (see Tetrahedron Letters (1981), 22, 1859).

In the PCR, gene amplification was performed by using DNA Thermal Cycler Model PJ2000 produced by Takara Shuzo Co., Ltd., using Taq DNA polymerase in accordance with a method designated by the supplier. After confirming an amplified gene fragment of 1643 kb by agarose gel electrophoresis, the fragment excised from the gel was purified in accordance with an ordinary method, and cut with restriction enzymes NruI (produced by Takara Shuzo Co., Ltd.) and EcoRI (produced by Takara Shuzo Co., Ltd.).

pHSG399 (see Takeshita, S. et al., Gene (1987), 61, 63-74) was used for a vector for cloning the gene fragment. pHSG399 was cut with a restriction enzyme SmaI (produced by Takara Shuzo Co., Ltd.) and a restriction enzyme EcoRI, and ligated with the amplified AK gene fragment. The ligation of DNA was performed in accordance with a designated method by using a DNA ligation kit (produced by Takara Shuzo Co., Ltd.). Thus a plasmid was prepared in which pHSG399 is connected to the AK gene product amplified from the chromosome of Brevibacterium. A plasmid having the AK gene originating from the 2256 strain (ATCC 13869) as a wild strain was designated as p399AKY, and a plasmid having the AK gene originating from AJ3463 as an L-lysine-producing bacterium was designated as p399AK9.

A DNA fragment having an ability to enable plasmids to make autonomous replication in bacteria belonging to the genus Corynebacterium (hereinafter referred to as "Coryne.-ori") was introduced into p399AKY and p399AK9 respectively, to prepare plasmids carrying the AK genes autonomously replicable in bacteria belonging to the genus Corynebacterium. Coryne.-ori was prepared from a plasmid vector autonomously replicable in bacterial cells of both Escherichia coli and bacteria belonging to the genus Corynebacterium. Some of such plasmid vectors have been reported. However, in this case, a shuttle vector pHK4 was used which was prepared from a plasmid pAJ1844 autonomously replicable in cells of coryneform bacteria (see Japanese Patent Laid-open No. 58-216199) and a plasmid pHSG298 autonomously replicable in cells of Escherichia coli (see Takeshita, S. et al., Gene, 61, 63-74 (1987)).

The preparation method for pHK4 is described in detail in Japanese Patent Laid-open No. 5-7491, however, it may be outlined as follows. pAJ1844 was partially cut with a restriction enzyme Sau3AI, and ligated with pHSG298 completely cut with a restriction enzyme BamHI. DNA after the ligation was introduced into Brevibacterium lactofermentum AJ12036 (FERM-P7559). An electric pulse method (see Japanese Patent Laid-open No. 2-207791) was used as a method for transformation. Selection of transformants was performed by using M-CM2G plates containing 25 µg/ml of kanamycin (containing 5 g of glucose, 10 g of polypeptone, 10 g of yeast extract, 5 g of NaCl, 0.2 g of DL-methionine and 15 g of agar in 1 l of pure water (pH 7.2)). Plasmids were prepared from transformants, and one having the smallest size was selected and designated as pHK4. This plasmid can make autonomous replication in Escherichia coli and coryneform bacteria, and gives kanamycin resistance to a host.

pHK4 obtained as described above was cut with a restriction enzyme KpnI (produced by Takara Shuzo Co., Ltd.), and cut faces were blunt-ended. Formation of blunt ends was performed in accordance with a designated method by using a DNA Blunting kit (produced by Takara Shuzo Co., Ltd.). After the blunt end formation, a phosphatized BamHI linker (produced by Takara Shuzo Co., Ltd.) was connected, to make modification to allow a DNA fragment of the Coryne.-ori portion to be cut from pHK4 with only BamHI. This plasmid was cut with BamHI. A generated Coryne.-ori DNA fragment was ligated with p399AKY or p399AK9 having been cut with BamHI in the same manner, to prepare a plasmid autonomously replicable in bacteria belonging to the genus Corynebacterium and contained the AK gene.

A plasmid containing the wild type AK gene originating from p399AKY was designated as p399AKYB, and a plasmid containing the mutant type AK gene originating from p399AK9 was designated as p399AK9B. Process of construc-

tion of p399AK9B and p399AKYB is shown in Fig. 4. A strain AJ12691 obtained by introducing the mutant type AK plasmid p399AK9B into the AJ12036 strain (FERM-P7559) as a wild type strain of Brevibacterium lactofermentum has been deposited on April 10, 1992 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology under a deposition number of FERM P-12918, transferred to international deposition based on the Budapest Treaty on February 10, 1995, and deposited under a deposition number of FERM BP-4999.

(2) Determination of nucleotide sequences of wild type and mutant type AK genes of Brevibacterium lactofermentum

The plasmid p399AKY containing the wild type AK gene and the plasmid p399AK9 containing the mutant type AK gene were prepared from each of the transformants, and nucleotide sequences of the wild and mutant type AK genes were determined. The determination of the nucleotide sequences was performed in accordance with a method of Sanger (F. Sanger et al., Proc. Natl. Acad. Sci., 74, 5463 (1977) and so on).

The nucleotide sequence of the wild type AK gene encoded by p399AKY is shown in SEQ ID NO: 7 in Sequence Listing. On the other hand, the nucleotide sequence of the mutant type AK gene encoded by p399AK9 only had mutation of one base pair in which 1051th G was changed to A in SEQ ID NO: 7 as compared with the wild type AK. It is known for the AK gene that two subunits of α , β are encoded on an identical DNA strand in an identical reading frame (see Kalinowski, J. et al., Molecular Microbiology (1991), 5(5), 1197-1204). Judging from homology, it is speculated for the gene of this case that two subunits of α , β are encoded on an identical DNA strand in an identical reading frame.

An amino acid sequence of the α -subunit of the wild type AK protein deduced from the nucleotide sequence of DNA is shown in SEQ ID NO: 8 in Sequence Listing simultaneously with the DNA sequence. Only the amino acid sequence is shown in SEQ ID NO: 9. An amino acid sequence of the β -subunit of the wild type AK protein deduced from the nucleotide sequence of DNA is shown in SEQ ID NO: 10 in Sequence Listing simultaneously with the DNA sequence. Only the amino acid sequence is shown in SEQ ID NO: 11. Each of the subunits uses GTG as a start codon, and a corresponding amino acid is represented as methionine. However, this represents methionine, valine or formylmethionine.

On the other hand, the mutation on the mutant type AK gene sequence indicates occurrence of replacement of amino acid residues such that a 279th alanine residue is replaced with a threonine residue in the α -subunit, and a 30th alanine residue is replaced with a threonine residue in the β -subunit, with respect to the amino acid sequences (SEQ ID NOS: 8 and 10) of the wild type AK protein.

(3) AK activity of expression product of mutant type AK gene and evaluation of desensitization of inhibition

Strains were prepared in which the wild type AK plasmid p399AKYB and the mutant type AK plasmid p399AK9B were respectively introduced into the AJ12036 strain (FERM-P7559) as a wild type strain of Brevibacterium lactofermentum (Corynebacterium glutamicum). The gene introduction into Corynebacterium was performed in accordance with an electric pulse method. The AK activity was measured for Brevibacterium lactofermentum (Corynebacterium glutamicum) AJ12036 strain as the host, an AJ12690 strain harboring the wild type AK plasmid, and an AJ12691 (FERM-P12918) strain harboring the mutant type AK plasmid. The measurement of the activity was performed in accordance with an ordinary method (see Miyajima, R. et al., The Journal of Biochemistry (1968), 63(2), 139-148).

As shown in Table 3, it has been confirmed that owing to the introduction of the AK plasmids, the specific activity of AK is increased about 10-15 times, and that the cumulative inhibition by L-lysine and L-threonine is desensitized only for the strain with the introduced mutant type AK plasmid. Table 3 shows the AK specific activity and the degree of its cumulative inhibition by L-lysine and L-threonine with respect to solutions obtained by destroying bacterial cells of the wild type AJ12036 strain of Brevibacterium lactofermentum, the AJ12690 strain allowed to harbor the wild type AK plasmid, and the AJ12691 strain allowed to harbor the mutant type AK plasmid. L-lysine and L-threonine as inhibitors were added to give a final concentration of 1 mM, respectively.

Table 3

Bacterial strain	AK specific activity (mU/mg protein)	
	No addition	+1mM L-lysine, +1mM L-threonine
AJ12036	19.0	2.6
AJ12690	235.3	34.6
AJ12691	210.5	145.3

(4) Improvement of mutant type AK gene by site-specific mutation

In order to further improve the mutant type AK obtained as described above, it was intended to replace the mutation point (²⁷⁹Ala → Thr) of the mutant type AK with another amino acid residue by means of site-specific mutation. The method for site-specific mutation for causing desired mutation at a desired site includes, for example, a method using PCR (Higuchi, R., 61, in *PCR Technology* (Erich, H. A. Eds., Stockton Press (1989))), a method using phage (Kramer, W. and Frits, H. J., *Meth. in Enzymol.*, 154, 350 (1987); Kunkel, T. A. et al., *Meth. in Enzymol.*, 154, 367 (1987)).

With respect to the species of amino acid residues to be introduced by mutation, 20 species of amino acids were classified in accordance with respective properties such as polarity and molecular structure, and representative 8 species (Arg, Asp, Cys, Phe, Pro, Ser, Tyr, Val) were selected. Amino acid mutation and nucleotide replacement at respective mutation points are shown in Table 4.

Table 4

Identification of mutation	Mutation point and amino acid change
Thr	²⁷⁹ Ala GCT → Thr A*CT
Arg	²⁷⁹ Ala GCT → Arg C*G*T
Asp	²⁷⁹ Ala GCT → Asp GA*T
Cys	²⁷⁹ Ala GCT → Cys T*G*T
Phe	²⁷⁹ Ala GCT → Phe T*T*T
Pro	²⁷⁹ Ala GCT → Pro C*CT
Ser	²⁷⁹ Ala GCT → Ser T*CT
Tyr	²⁷⁹ Ala GCT → Tyr T*A*T
Val	²⁷⁹ Ala GCT → Val GT*T

A method for introducing mutation used herein is as follows. Eight species of synthetic DNA of 23 mers, in which the codon for the 279th Ala residue for introducing mutation was replaced with codons for desired amino acid residues, were designed (5'-GCCAGGCGAG CGT GCCAAGGTTT-3': SEQ ID NO: 12 as synthetic DNA for introducing Arg; 5'-GCCAGGCGAG GAT GCCAAGGTTT-3': SEQ ID NO: 13 as synthetic DNA for introducing Asp; 5'-GCCAGGCGAG TGT GCCAAGGTTT-3': SEQ ID NO: 14 as synthetic DNA for introducing Cys; 5'-GCCAGGCGAG TTT GCCAAGGTTT-3': SEQ ID NO: 15 as synthetic DNA for introducing Phe; 5'-GCCAGGCGAG CCT GCCAAGGTTT-3': SEQ ID NO: 16 as synthetic DNA for introducing Pro; 5'-GCCAGGCGAG TCT GCCAAGGTTT-3': SEQ ID NO: 17 as synthetic DNA for introducing Ser; 5'-GCCAGGCGAG TAT GCCAAGGTTT-3': SEQ ID NO: 18 as synthetic DNA for introducing Tyr; and 5'-GCCAGGCGAG GTT GCCAAGGTTT-3': SEQ ID NO: 19 as synthetic DNA for introducing Val). Sixteen species of 23 mer single strand DNA's were synthesized together with their complementary sequences.

When an Arg residue is introduced, for example, the single strand DNA having the sequence 5'-GCCAGGCGAG CGT GCCAAGGTTT-3' (SEQ ID NO: 12), the single strand DNA as its complementary chain, the single strand DNA having the sequence of SEQ ID NO: 5, and the single strand DNA having the sequence of SEQ ID NO: 6 were used as primers, and the PCR method was performed using p399AKY as a template. In order to avoid introduction of nonspecific mutation, about 280 base pairs containing the mutation point were excised from prepared DNA with restriction enzymes (*NaeI*-*AvaII*), and replaced with a corresponding site of p399AKY to prepare a recombinant plasmid. The nucleotide sequence was confirmed for the replaced region.

Upon measurement and evaluation of the enzyme activities of the mutant type AK's harbored by each of 8 species of obtained recombinant plasmids, an AK completely deficient strain of *E. coli*, Gif106M1 was used as a host (Boy, E. and Patte, J. C., *J. Bacteriol.*, 112, 84-92 (1972); Theze, J. et al., *J. Bacteriol.*, 117, 133-143 (1974)), because no AK deficient strain was known for coryneform bacteria. Otherwise, AK of a host and AK from the plasmid may exist in a mixed manner, probably resulting in inaccurate measurement. Many genes of coryneform bacteria are known to be expressed in *E. coli*. Thus it was postulated that the AK gene could be expressed in *Escherichia coli* since it was linked downstream from a *lac* promoter on pHSG399.

E. coli Gif106M1 was transformed with the recombinant plasmids of the wild type and the eight species, cell-free

extracts were prepared from each of transformed strains, and analysis of enzyme was performed. The AK activity was measured in accordance with a method described in Miyajima, R. et al., *The Journal of Biochemistry* (1968), 63(2), 139-148. The degree of inhibition desensitization and the specific activity are shown in Table 5, in the case of addition of 5 mM of L-lysine, 5 mM of L-threonine, or each 2 mM of L-lysine and L-threonine.

Table 5

	Specific activity (mU/mg protein)	5 mM Lys (%)	5 mM Thr (%)	2 mM Lys + Thr (%)
AJ12036	5.6	52.0	87.0	7.0
Wild type	316.4	52.7	86.8	6.2
Thr	374.4	58.7	109.1	78.3
Arg	197.4	41.4	106.8	58.6
Cys	267.0	66.5	135.7	60.6
Phe	447.7	14.6	105.0	32.4
Pro	125.0	77.5	123.2	85.2
Ser	406.8	55.0	114.4	37.0
Tyr	425.6	16.1	104.8	32.2
Val	448.9	60.5	103.5	75.5

As a result, AK was inactivated in the case of change to acidic amino acid such as Asp, while the inhibition by L-lysine and L-threonine was desensitized in the case of change to any other amino acid.

Example 3: Evaluation of L-lysine Productivity of HD Mutant Strain and HD Deficient Strain

In order to compare effects on the L-lysine productivity exerted by the two kinds of mutant type HD and the HD deficiency, mutant type HD genes or an HD gene with a part of its sequence deleted was integrated into chromosome of an identical host to prepare gene-replaced strains each of which was used as HD mutant strains and an HD deficient strain, and evaluated for the L-lysine productivity.

(1) Preparation of plasmids for replacing mutant type HD genes and plasmid for replacing deficient type HD gene

Plasmids for gene replacement were prepared for introduction by homologous recombination of mutant type HD genes or an HD gene with a part of its sequence deleted, into chromosomal DNA of *Brevibacterium lactofermentum* AJ12036 strain (FERM BP-734) (obtained by deleting a cryptic plasmid, pAM330 from *Brevibacterium lactofermentum* 2256 strain (ATCC 13869)).

(1) Preparation of HD gene having mutation point 1

The mutant type HD genes obtained in Example 1 were two species including the mutant type HD gene (from AJ12472 strain) having the mutation point 1 ($^{155}\text{C} \rightarrow \text{T}$ ($^{23}\text{Leu} \rightarrow \text{Phe}$)) and the mutation point 2 ($^{398}\text{G} \rightarrow \text{A}$ ($^{104}\text{Val} \rightarrow \text{Ile}$)), and the mutant type gene (from AJ12937) having only the mutation point 2. In order to investigate the influence exerted by the mutation point 1 on the HD activity and the L-lysine productivity, a mutant type HD gene having only the mutation point 1 was prepared. Hereinafter, the mutant type HD having the mutation point 1 is referred to as HD-M1, the mutant type HD having the mutation point 2 is referred to as HD-M2, and the mutant type HD having both the mutation points 1 and 2 is referred to as HDM-12.

The plasmid pHDMI containing the HDM-12 gene was cut with a restriction enzyme *TthIII1* for cutting between the both mutation points of 1 and 2, and *KpnI* for cutting a ligation point between the vector and the HD gene, to obtain a 5' side HD fragment having the mutation point 1. In the same manner, pHDW having the wild type HD gene was cut with *TthIII1* and *KpnI* to obtain a 3' side HD fragment. The HD-M1 gene having only the mutation point 1 was obtained by ligating the 5' side HD fragment with the 3' side HD fragment thus obtained.

(2) Construction of plasmids for gene replacement

The HD-M1 gene having only the mutation point 1 obtained as described above was inserted into a KpnI site of a vector plasmid pHSG398 having a chloramphenicol resistance (Cm^r) gene. Further, a temperature-sensitive replication origin (TSori) originating from Brevibacterium lactofermentum wild strain was introduced into a BamHI site of pHSG398. Thus a plasmid pTSHDM1 for replacing the HD-M1 gene was constructed. TSori was prepared from a plasmid pHSC4 (see Japanese Patent Laid-open No. 5-7491) obtained by treating a plasmid pHK4 having Coryne.-ori with hydroxylamine in vitro, transforming Brevibacterium lactofermentum AJ12036 with plasmid DNA after the treatment, and recovering the plasmid from a transformed strain incapable of growth at a high temperature (34°C). Coryne.-ori can be excised from pHSC4 with BamHI and KpnI, however, the plasmid was modified to allow Coryne.-ori to be excised only by cutting with BamHI. pHSC4 was cut with a restriction enzyme KpnI (produced by Takara Shuzo Co., Ltd.), and cut faces were blunt-ended. Formation of blunt ends was performed in accordance with a designated method using a DNA Blunting kit (produced by Takara Shuzo Co., Ltd.). After the blunt end formation, a phosphatized BamHI linker (produced by Takara Shuzo Co., Ltd.) was connected, to make modification to allow a DNA fragment of the TSori portion to be excised from pHSC4 with only BamHI. Escherichia coli AJ12571 harboring pHSC4 has been deposited on October 11, 1990 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology under a deposition number of FERM P-11763, transferred to international deposition based on the Budapest Treaty on August 26, 1991, and deposited under a deposition number of FERM BP-3524.

Next, in the same manner, the plasmid pHDII having the HD-M2 gene having only the mutation point 2 was cut with KpnI to obtain an HD-M2 gene fragment which was inserted into a KpnI site of pHSG398. Subsequently, TSori was inserted into a BamHI site. Thus a plasmid pTSHDM2 for replacing the HD-M2 gene was constructed.

Further, the plasmid pHDMI having the HD-M12 gene having both the mutation points 1 and 2 was cut with KpnI to obtain an HD-M12 gene fragment which was inserted into a KpnI site of pHSG398. Subsequently, TSori was inserted into a BamHI site. Thus a plasmid pTSHDM12 for replacing the HD-M12 gene was constructed.

Further, the plasmid pHDW having the wild type HD gene was cut with AatII, and a portion between two AatII sites (nucleotide numbers of 716-722 and 1082-1087 in SEQ ID NO: 3) existing in the HD gene was deleted. Thus a plasmid containing an HD gene with its part deleted (HD-Δ gene) was prepared. This plasmid was cut with KpnI to obtain an HD-Δ gene fragment which was inserted into a KpnI site of pHSG398. Subsequently TSori was inserted into a BamHI site. Thus a plasmid pTSHDA for replacing the HD-Δ gene was constructed.

(3) Preparation of HD mutant strain and HD deficient strain

Transformation of Brevibacterium lactofermentum AJ12036 strain was performed in accordance with an electric pulse method (Sugimoto et al., Japanese Patent Laid-open No. 2-207791) by using the plasmids for replacing the mutant type HD genes pTSHDM1, pTSHDM2 and pTSHDM12, and the plasmid for replacing the deletion type HD gene pTSHDA obtained as described above.

Obtained transformed strains were cultivated by using an M-CM2G medium at 25°C until full growth (about 1-2 x 10⁹/ml) was achieved. Cultivated bacterial cells were diluted to give 10⁵ cells per one plate, spread on an M-CM2G solid plate medium containing chloramphenicol (5 μg/mL), and cultivated at 34°C for 2-7 days to obtain colonies. It was confirmed for the obtained colonies that no plasmid was contained in cells. Further, it was confirmed that the plasmids for gene replacement were integrated into chromosome by means of Southern hybridization analysis using linear pHSG398 as a probe.

In the strain of chromosomal integration obtained as described above, two fused genes of an HD gene originally existing on the chromosome and the mutant type or deletion type HD gene are inserted in a state of interposing the vector (including TSori).

Next, in order to leave only the mutant type HD gene or the deletion type HD gene on the chromosome, the wild type HD gene and the vector were allowed to fall off from the chromosomal DNA to obtain strains replaced with the mutant type HD genes and a strain replaced with the deletion type HD gene. The wild type HD gene and the vector were allowed to fall off as follows.

Each of the integrated strains was cultivated at 34°C in an M-CM2G medium containing chloramphenicol (10 μg/mL) until full growth (1-2 x 10⁹) was achieved. Cultivated bacterial cells were spread on an M-CM2G solid plate medium containing no chloramphenicol to give 50-200 colonies per one plate, and cultivated at 34°C. Grown colonies were replicated onto an M-CM2G solid plate medium containing chloramphenicol (5 μg/mL), and cultivated at 34°C to obtain chloramphenicol sensitive strains. It was confirmed by Southern hybridization that the vector fell off from chromosome of the obtained chloramphenicol sensitive strains. It was further confirmed that the mutant type HD or the deletion type HD was expressed. It was confirmed by nucleotide sequence determination of chromosomal DNA that the mutation points were introduced into the gene-replaced strains thus obtained.

The HD-M1 gene-replaced strain thus obtained is designated as HDM1 strain, the HD-M2 gene-replaced strain is designated as HDM2 strain, the HDM-12 gene-replaced strain is designated as HDM12 strain, and the HD-Δ gene-

replaced strain is designated as HDA strain.

(2) L-lysine productivity of HD mutant strain and HD deficient strain

The L-lysine productivity was investigated for the HDM1, HDM2 and HDM12 strains as the HD mutant strains, and for the HDA strain as the HD deficient strain. These HD mutant strains and HD deficient strain purified by single colony isolation, as well as the AJ12036 strain as the wild strain for HD were respectively cultivated in a flask of 500 mL added with 20 mL of an L-lysine production medium shown below at 31.5°C for 72 hours. The final OD (OD₅₆₂) and accumulated amount of L-lysine were examined.

(L-lysine production medium)

This medium was prepared by dissolving components described below (in 1 L), adjusting pH to 8.0 with KOH, sterilizing at 115°C for 15 minutes, and then adding 50 g/L of CaCO₃ having been sterilized by heat in a dry state.

Glucose	100 g
(NH ₄) ₂ SO ₄	55 g
KH ₂ PO ₄	1 g
MgSO ₄ · 7H ₂ O	1 g
d-biotin	500 µg
Thiamin-HCl	2000 µg
FeSO ₄ · 7H ₂ O	0.01 g
MnSO ₄ · 7H ₂ O	0.01 g
Nicotinamide	5 mg
Mamenou (T-N)	1.05 g
GD113	0.05 ml

Results are shown in Fig. 5. No remaining sugar was found in any strain. As clarified from the results, accumulation of L-lysine was scarcely observed in AJ12036 strain, while it was about 4 g/l for HDM1 strain, about 17 g/l for HDM2 strain, about 7.5 g/l for HDM12 strain, and about 30 g/l for HDA strain. Accumulation of L-lysine was observed in any of the latter strains. Especially the L-lysine productivity was remarkably improved in HDA strain. Further, it was clarified that L-lysine was accumulated by introduction of only the mutation point 1 into HD.

HDA strain did not grow in a minimum medium or another minimum medium added with L-threonine or L-methionine alone, however, its growth was recovered by addition of L-homoserine, or L-threonine and L-methionine. Any of HDM1, HDM2 and HDM12 strains could grow in a minimum medium containing neither L-threonine nor L-methionine.

Brevibacterium lactofermentum HDA strain was designated as Brevibacterium lactofermentum AJ12846. It has been deposited on March 1, 1994 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology under a deposition number of FERM P-14197, transferred to international deposition based on the Budapest Treaty on February 9, 1995, and deposited under a deposition number of FERM BP-4995.

Example 4: Effect of Amplification of AK Gene in HD Mutant Strains and HD Deficient Strain

As described in Example 3, it has been clarified that the L-lysine productivity is improved by introducing the mutant type HD genes and the deletion type HD gene into the wild strain. Further investigation was made for the effect expected by combining AK gene amplification with the mutant type HD genes and the deletion type HD gene.

It is known that AK undergoes concerted inhibition by L-lysine and L-threonine, however, the degree of inhibition is low in the case of the presence of each of them alone. Therefore, since no L-threonine is produced by the HD mutant strain and the HD deficient strain, it is expected that the L-lysine productivity may be also improved by amplification of the wild type AK gene. It is further expected that the L-lysine productivity may be more improved by introducing the gene coding for the mutant type AK obtained in Example 2 which does not undergo inhibition by L-lysine and L-threonine.

In order to investigate such a combined effect of introduction of the mutant type HD gene or the deletion type HD gene and amplification of the AK gene, plasmids containing the AK genes were introduced into the HD mutant strains

and the HD deficient strain obtained in Example 3, and the L-lysine productivity was evaluated.

Each of the strains of AJ12036 as a wild strain, HDM1, HDM2 and HDM12 as HD mutant strains, and HDA as an HD deficient strain was used as hosts respectively, and transformed with the plasmid (p399AKYB) having the wild type AK gene and Coryne.-ori and the plasmid (p399AK9B) having the mutant type AK. Namely, 5 species of the hosts were transformed with 2 species of the plasmids, and 10 species of transformed strains were obtained in total.

Two strains for each of the transformed strains of AJ12036, HDM1, HDM2, HDM12 and HDA were cultivated by using the aforementioned L-lysine production medium, and the L-lysine productivity was examined. However, the transformed strains harboring the p399AKYB plasmid and the p399AK9B plasmid were cultivated with addition of 10 µg/mL of chloramphenicol to both a medium used for pre-cultivation and the L-lysine production medium. The cultivation was performed with stirring at 31.5°C for 72 hours in a flask of 500 mL added with 20 mL of the medium.

According to the result as shown in Fig. 6, no improvement in L-lysine productivity was observed for the AJ12036 strain even when the wild type AK plasmid was introduced, while increase in accumulated amount of L-lysine was observed for the HD mutant strain and the HD deficient strain owing to the introduction of the wild type AK plasmid. Further, when the mutant type AK plasmid was introduced, the L-lysine productivity was more improved for any of the HD mutant strains and the HD deficient strain as compared with the case of the introduction of the wild type AK plasmid. Furthermore, accumulation of L-lysine in an amount of about 22 g/L was observed even in the case of the AJ12036 strain harboring the wild type HD gene owing to the introduction of the mutant type AK plasmid.

Example 5: Effect of Gene Replacement of Mutant Type AK Gene in HD Mutant Strains and HD Deficient Strain

(1) Creation of strains replaced with mutant type AK gene and mutant type HD genes, and strain replaced with mutant type AK and deletion type HD gene

The effect of combination of HD mutation and HD deficiency with AK gene amplification has been investigated in Example 4. This Example concerns strains created such that the mutant type HD gene or the deletion type HD gene is integrated on chromosome, and the mutant type AK gene is integrated on chromosome, in order to evaluate the L-lysine productivity.

A plasmid for gene replacement for integrating the mutant type AK gene into chromosomal DNA was obtained as follows.

A plasmid pAK9T for replacing the mutant type AK gene was constructed by inserting the temperature-sensitive replication origin (TSori) of *Brevibacterium lactofermentum* into a *Bam*HI site existing at a vector portion of the plasmid p399AK9 obtained in Example 2 (plasmid with the mutant type AK gene fragment originating from *Brevibacterium lactofermentum* AJ3463 strain amplified from chromosome connected to pHSG399).

A strain introduced with the mutant type AK gene and the HD-M1 gene [(AK^{FBR}+HDM1) strain] was obtained by integrating the mutant type AK gene by employing the plasmid for replacing the mutant type AK gene pAK9T, using the HDM1 strain obtained in Example 4 as a parent strain. pAK9T was introduced into the HDM1 strain by means of an electric pulse method (Sugimoto et al., Japanese Patent Laid-open No. 2-207791), and obtained transformed strains were cultivated at 25°C by using an M-CM2G medium until full growth (about 1-2 x 10⁹/ml) was achieved. Cultivated bacterial cells were diluted to give 10⁵ cells per one plate, spread on an M-CM2G solid plate medium containing chloramphenicol (5 µg/mL), and cultivated at 34°C for 2-7 days to obtain colonies. It was confirmed for obtained colonies that no plasmid was contained in cells. Further, integration of pAK9T into chromosome was confirmed by Southern hybridization analysis using linear pHSG399 as a probe.

In the strain of chromosomal integration obtained as described above, two fused genes of an AK gene originally existing on the chromosome and the mutant type AK gene are inserted into chromosome in a state of interposing the vector (including TSori). Next, in order to leave only the mutant type AK gene on chromosomal DNA, the wild type AK gene and the vector were allowed to fall off to obtain a strain replaced with the mutant type AK gene. The vector was allowed to fall off as follows.

The strain integrated with the mutant type AK gene was cultivated in an M-CM2G medium containing chloramphenicol (10 µg/mL) at 34°C until full growth (1-2 x 10⁹) was achieved. Cultivated bacterial cells were spread on an M-CM2G solid plate medium containing no chloramphenicol to give 50-200 colonies per one plate, and cultivated at 34°C. Among clones which formed colonies, a strain was selected in which the L-lysine productivity was improved as compared with the HDM1 strain as the parent strain.

In the same manner, a strain having the mutant type AK gene and the HD-M12 gene [(AK^{FBR}+HDM12) strain] was selected as a strain with improved L-lysine productivity compared with the HDM12 strain, by performing gene replacement employing pAK9T in the same manner as described above using the HDM12 strain as a parent strain.

On the other hand, a strain introduced with the mutant type AK gene and the HD-M2 gene [(AK^{FBR}+HDM2) strain] was obtained by creating an AK^{FBR} strain having the introduced mutant type AK gene as a parent strain by introducing pAK9T into *Brevibacterium lactofermentum* AJ12036 strain, followed by introduction of the HD-M2 gene. Namely, pAK9T was introduced into the AJ12036 strain by means of an electric pulse method (Sugimoto et al., Japanese Patent

Laid-open No. 2-207791), and obtained transformed strains were cultivated at 25°C by using an M-CM2G medium until full growth (about $1-2 \times 10^9$ /ml) was achieved. Cultivated bacterial cells were diluted to give 10^5 cells per one plate, spread on an M-CM2G solid plate medium containing chloramphenicol (5 µg/mL), and cultivated at 34°C for 2-7 days to obtain colonies. It was confirmed for obtained colonies that no plasmid was contained in cells. Further, integration of pAK9T into chromosome was confirmed by Southern hybridization analysis using linear pHSG399 as a probe.

Next, a strain integrated with the mutant type AK gene was cultivated in an M-CM2G medium containing chloramphenicol (10 µg/mL) at 34°C until full growth ($1-2 \times 10^9$) was achieved. Cultivated bacterial cells were spread on an M-CM2G solid plate medium containing no chloramphenicol to give 50-200 colonies per one plate, and cultivated at 34°C. Grown colonies were replicated onto an M-CM2G solid plate medium containing chloramphenicol (5 µg/mL), and cultivated at 34°C to obtain a chloramphenicol sensitive strain. It was confirmed by Southern hybridization that the vector fell off from chromosome of the obtained chloramphenicol sensitive strain. It was further confirmed that the mutant type AK was expressed. It was confirmed by nucleotide sequence determination of chromosomal DNA that the gene-replaced strain thus obtained had the introduced mutation point.

The plasmid pTSHDM2 for replacing the HD-M2 gene was introduced into chromosome of the mutant type AK gene-replaced strain (AK^{FBR} strain) thus obtained, by means of the electric pulse method in the same manner as described above, and a gene-replaced strain in which the plasmid fell off was obtained. The gene-replaced strain was selected in accordance with the improvement in L-lysine productivity and sensitivity to L-threonine or L-methionine.

In the same manner, a strain introduced with the mutant type AK gene and the HD-Δ gene [$(AK^{FBR}+HD\Delta)$] was obtained by using the mutant type AK gene-replaced strain (AK^{FBR} strain) as a parent strain, performing gene replacement with pTSHDΔ in the same manner as described above, and selecting a clone auxotrophic for L-methionine and L-threonine due to HD deficiency.

For each of the gene-replaced strains obtained as described above, it was finally confirmed by nucleic acid sequence determination of DNA that mutation points were introduced. Further, it was confirmed by Southern hybridization that the plasmid fell off.

(2) Evaluation of L-lysine productivity of strains replaced with mutant type AK gene and mutant type HD genes, and strain replaced with mutant type AK gene and deletion type HD gene

The L-lysine productivity was evaluated for the four strains obtained as described above, namely, ($AK^{FBR}+HDM1$), ($AK^{FBR}+HDM2$), ($AK^{FBR}+HDM12$) and ($AK^{FBR}+HD\Delta$) strains.

Each of these strains was cultivated at 31.5°C for 72 hours in a flask of 500 mL added with 20 mL of the aforementioned L-lysine production medium, to measure OD of culture liquid and the amount of accumulated L-lysine after the cultivation.

Results are shown in Fig. 7. The amount of produced L-lysine was about 19 g/L for the AK^{FBR} strain, while it was about 21 g/L for the ($AK^{FBR}+HDM1$) strain, about 22 g/L for the ($AK^{FBR}+HDM2$) strain, about 20 g/L for the ($AK^{FBR}+HDM12$) strain, and about 35 g/L for the ($AK^{FBR}+HD\Delta$) strain. It was indicated that the L-lysine productivity was more improved when the mutant type HD genes or the deletion type HD gene was introduced in combination with the mutant type AK gene than when they were introduced alone.

There was little difference in OD of media after completion of the cultivation between the AK^{FBR} strain and any of the ($AK^{FBR}+HDM1$), ($AK^{FBR}+HDM2$) and ($AK^{FBR}+HDM12$) strains. However, OD was more decreased in the case of the ($AK^{FBR}+HD\Delta$) strain as compared with the gene-replaced strain with the deletion type HD gene alone. No remaining sugar was found after completion of the cultivation in any of the strains.

The ($AK^{FBR}+HDM2$), ($AK^{FBR}+HDM12$) and ($AK^{FBR}+HD\Delta$) strains were designated as AJ12848 (FERM P-14198), AJ12849 (FERM P-14199) and AJ12850 (FERM P-14200), respectively. They have been deposited on March 1, 1994 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology under the deposition numbers described above in the parentheses respectively, transferred to international deposition based on the Budapest Treaty on February 9, 1995, and deposited under deposition numbers of FERM BP-4996, FERM BP-4997 and FERM BP-4998 respectively in this order.

Example 6: Measurement of Reverse Mutation Frequency of HD Completely Deficient Strain

The reverse mutation frequency for homoserine auxotrophy was compared between the completely deficient strains of HD (HDA and $AK^{FBR}+HDA$ strains) obtained by the gene replacement on chromosome and an HD deficient strain ATCC 13287 obtained by treating living bacterial cells with N-methyl-N'-nitro-N-nitrosoguanidine (NTG) as an ordinary mutating treatment agent.

The method for comparison was as follows. Stocked bacterial strains were pre-cultivated in a nutrient-rich medium, and inoculated in an L-lysine production medium. After cultivation with stirring for 72 hours, the culture liquids were appropriately diluted. Colonies were formed on an M-CM2G solid plate medium, which were subsequently replicated onto a minimum medium for *Brevibacterium* containing neither L-methionine nor L-threonine. The ratio of colonies

EP 0 754 756 A1

grown on the minimum medium to those grown on the nutrient-rich medium was regarded as a ratio of reverse mutation. When no revertant strain could be observed by using this method, the number of bacterial cells was increased to be applied to the minimum medium, and the number of bacterial cells was changed in accordance with degrees of dilution to be applied to the nutrient-rich medium. The number of bacterial cells applied to the minimum medium was estimated, and the ratio was calculated.

Results of measurement of the reverse mutation frequencies of the aforementioned three strains by means of this method are shown in Table 6. Revertant strains considerably appeared upon completion of the cultivation in the case of the ATCC 13287 strain. On the contrary, no revertant strain was observed at all in the case of the HDA and AK^{FBR}+HDA strains prepared by chromosomal recombination. Further, the HDA strain with no occurrence of reverse mutation had a larger amount of produced L-lysine than the ATCC 13287 strain.

Table 6

Bacterial strain	Reverse mutation frequency (%)	Accumulated amount of L-lysine (g/l)
ATCC 13287	40	20.0
HDA	0	30.0
AK ^{FBR} +HDA	0	35.0

EP 0 754 756 A1

SEQUENCE LISTING

INFORMATION FOR SEQ ID NO:1:

SEQUENCE CHARACTERISTICS:

LENGTH: 20

TYPE: nucleic acid

STRANDEDNESS: single

TOPOLOGY: linear

MOLECULAR TYPE: other..synthetic oligonucleotide

SEQUENCE DESCRIPTION: SEQ ID NO:1:

CTGGGAAGGT GAATCGAATT

INFORMATION FOR SEQ ID NO:2:

SEQUENCE CHARACTERISTICS:

LENGTH: 20

TYPE: nucleic acid

STRANDEDNESS: single

TOPOLOGY: linear

MOLECULAR TYPE: other..synthetic oligonucleotide

SEQUENCE DESCRIPTION: SEQ ID NO:2:

TCCGAGGTTT GCAGAAGATC 20

INFORMATION FOR SEQ ID NO:3:

SEQUENCE CHARACTERISTICS:

LENGTH: 1478

TYPE: nucleic acid

STRANDEDNESS: double

TOPOLOGY: linear

MOLECULAR TYPE: genomic DNA

ORIGINAL SOURCE:

ORGANISM: Brevibacterium lactofermentum

STRAIN: AJ12036

FEATURE:

NAME/KEY: CDS

LOCATION: 89..1423

IDENTIFICATION METHOD: S

SEQUENCE DESCRIPTION: SEQ ID NO:3:

GGTACCCTTT TTGTTTGGGA CACATGTAGG GTGGCCGAAA CAAAGTAATA GGACAACAAC	60
GCTCGACCGC GATTATTTTT GGAGAATC ATG ACC TCA GCA TCT GCC CCA AGC	112
Met Thr Ser Ala Ser Ala Pro Ser	
1 5	
TTT AAC CCC GGC AAG GGT CCC GGC TCA GCA GTC GGA ATT GCC CTT TTA	160
Phe Asn Pro Gly Lys Gly Pro Gly Ser Ala Val Gly Ile Ala Leu Leu	
10 15 20	
GGA TTC GGA ACA GTC GGC ACT GAG GTG ATG CGT CTG ATG ACC GAG TAC	208
Gly Phe Gly Thr Val Gly Thr Glu Val Met Arg Leu Met Thr Glu Tyr	
25 30 35 40	
GGT GAT GAA CTT GCG CAC CGC ATT GGT GGC CCA CTG GAG GTT CGT GGC	256
Gly Asp Glu Leu Ala His Arg Ile Gly Gly Pro Leu Glu Val Arg Gly	
45 50 55	
ATT GCT GTT TCT GAT ATC TCA AAG CCA CGT GAA GGC GTT GCA CCT GAG	304
Ile Ala Val Ser Asp Ile Ser Lys Pro Arg Glu Gly Val Ala Pro Glu	
60 65 70	
CTG CTC ACT GAG GAC GCT TTT GCA CTC ATC GAG CGC GAG GAT GTT GAC	352
Leu Leu Thr Glu Asp Ala Phe Ala Leu Ile Glu Arg Glu Asp Val Asp	
75 80 85	
ATC GTC GTT GAG GTT ATC GGC GGC ATT GAG TAC CCA CGT GAG GTA GTT	400
Ile Val Val Glu Val Ile Gly Gly Ile Glu Tyr Pro Arg Glu Val Val	
90 95 100	

EP 0 754 756 A1

	CTC	GCA	GCT	CTG	AAG	GCC	GGC	AAG	TCT	GTT	GTT	ACC	GCC	AAT	AAG	GCT	448
	Leu	Ala	Ala	Leu	Lys	Ala	Gly	Lys	Ser	Val	Val	Thr	Ala	Asn	Lys	Ala	
	105					110					115					120	
5	CTT	GTT	GCA	GCT	CAC	TCT	GCT	GAG	CTT	GCT	GAT	GCA	GCG	GAA	GCC	GCA	496
	Leu	Val	Ala	Ala	His	Ser	Ala	Glu	Leu	Ala	Asp	Ala	Ala	Glu	Ala	Ala	
					125					130					135		
	AAC	GTT	GAC	CTG	TAC	TTC	GAG	GCT	GCT	GTT	GCA	GCC	GCA	ATT	CCA	GTG	544
	Asn	Val	Asp	Leu	Tyr	Phe	Glu	Ala	Ala	Val	Ala	Ala	Ala	Ile	Pro	Val	
				140						145				150			
10	GTT	GGC	CCA	CTG	CGT	CGC	TCC	CTG	GCT	GGC	GAT	CAG	ATC	CAG	TCT	GTG	592
	Val	Gly	Pro	Leu	Arg	Arg	Ser	Leu	Ala	Gly	Asp	Gln	Ile	Gln	Ser	Val	
		155						160				165					
	ATG	GGC	ATC	GTT	AAC	GGC	ACC	ACC	AAC	TTC	ATC	TTG	GAC	GCC	ATG	GAT	640
	Met	Gly	Ile	Val	Asn	Gly	Thr	Thr	Asn	Phe	Ile	Leu	Asp	Ala	Met	Asp	
		170					175				180						
15	TCC	ACC	GGC	GCT	GAC	TAT	GCA	GAT	TCT	TTG	GCT	GAG	GCA	ACT	CGT	TTG	688
	Ser	Thr	Gly	Ala	Asp	Tyr	Ala	Asp	Ser	Leu	Ala	Glu	Ala	Thr	Arg	Leu	
		185				190					195				200		
	GGT	TAC	GCC	GAA	GCT	GAT	CCA	ACT	GCA	GAC	GTC	GAA	GGC	CAT	GAC	GCC	736
	Gly	Tyr	Ala	Glu	Ala	Asp	Pro	Thr	Ala	Asp	Val	Glu	Gly	His	Asp	Ala	
				205						210				215			
20	GCA	TCC	AAG	GCT	GCA	ATT	TTG	GCA	TCC	ATC	GCT	TTC	CAC	ACC	CGT	GTT	784
	Ala	Ser	Lys	Ala	Ala	Ile	Leu	Ala	Ser	Ile	Ala	Phe	His	Thr	Arg	Val	
				220					225				230				
	ACC	GCG	GAT	GAT	GTG	TAC	TGC	GAA	GGT	ATC	AGC	AAC	ATC	AGC	GCT	GCC	832
	Thr	Ala	Asp	Asp	Val	Tyr	Cys	Glu	Gly	Ile	Ser	Asn	Ile	Ser	Ala	Ala	
25			235				240					245					
	GAC	ATT	GAG	GCA	GCA	CAG	CAG	GCA	GGC	CAC	ACC	ATC	AAG	TTG	TTG	GCC	880
	Asp	Ile	Glu	Ala	Ala	Gln	Gln	Ala	Gly	His	Thr	Ile	Lys	Leu	Leu	Ala	
		250				255					260						
	ATC	TGT	GAG	AAG	TTC	ACC	AAC	AAG	GAA	GGA	AAG	TCG	GCT	ATT	TCT	GCT	928
	Ile	Cys	Glu	Lys	Phe	Thr	Asn	Lys	Glu	Gly	Lys	Ser	Ala	Ile	Ser	Ala	
30		265			270					275					280		
	CGC	GTG	CAC	CCG	ACT	CTA	TTA	CCT	GTG	TCC	CAC	CCA	CTG	GCG	TCG	GTA	976
	Arg	Val	His	Pro	Thr	Leu	Leu	Pro	Val	Ser	His	Pro	Leu	Ala	Ser	Val	
				285					290				295				
	AAC	AAG	TCC	TTT	AAT	GCA	ATC	TTT	GTT	GAA	GCA	GAA	GCA	GCT	GGT	CGC	1024
35	Asn	Lys	Ser	Phe	Asn	Ala	Ile	Phe	Val	Glu	Ala	Glu	Ala	Ala	Gly	Arg	
				300					305				310				
	CTG	ATG	TTC	TAC	GGA	AAC	GGT	GCA	GGT	GGC	GCG	CCA	ACC	GCG	TCT	GCT	1072
	Leu	Met	Phe	Tyr	Gly	Asn	Gly	Ala	Gly	Gly	Ala	Pro	Thr	Ala	Ser	Ala	
			315				320					325					
	GTG	CTT	GGC	GAC	GTC	GTT	GGT	GCC	GCA	CGA	AAC	AAG	GTG	CAC	GGT	GGC	1120
40	Val	Leu	Gly	Asp	Val	Val	Gly	Ala	Ala	Arg	Asn	Lys	Val	His	Gly	Gly	
		330					335					340					
	CGT	GCT	CCA	GGT	GAG	TCC	ACC	TAC	GCT	AAC	CTG	CCG	ATC	GCT	GAT	TTC	1168
	Arg	Ala	Pro	Gly	Glu	Ser	Thr	Tyr	Ala	Asn	Leu	Pro	Ile	Ala	Asp	Phe	
		345				350					355				360		
	GGT	GAG	ACC	ACC	ACT	CGT	TAC	CAC	CTC	GAC	ATG	GAT	GTG	GAA	GAT	CGC	1216
45	Gly	Glu	Thr	Thr	Thr	Arg	Tyr	His	Leu	Asp	Met	Asp	Val	Glu	Asp	Arg	
				365						370					375		
	GTG	GGC	GTT	TTG	GCT	GAA	TTG	GCT	AGC	CTG	TTC	TCT	GAG	CAA	GGA	ATC	1264
	Val	Gly	Val	Leu	Ala	Glu	Leu	Ala	Ser	Leu	Phe	Ser	Glu	Gln	Gly	Ile	
				380					385				390				
50	TCC	CTG	CGT	ACA	ATC	CGA	CAG	GAA	GAG	CGC	GAT	GAT	GAT	GCA	CGT	CTG	1312
	Ser	Leu	Arg	Thr	Ile	Arg	Gln	Glu	Glu	Arg	Asp	Asp	Asp	Ala	Arg	Leu	
			395				400						405				

55

EP 0 754 756 A1

ATC GTT GTC ACG CAC TCT GCG CTG GAA TCT GAT CTT TCC CGC ACC GTT 1360
 Ile Val Val Thr His Ser Ala Leu Glu Ser Asp Leu Ser Arg Thr Val
 410 415 420
 5 GAA CTG CTG AAG GCT AAG CCT GTT GTT AAG GCA ATC AAC AGT GTG ATC 1408
 Glu Leu Leu Lys Ala Lys Pro Val Val Lys Ala Ile Asn Ser Val Ile
 425 430 435 440
 CGC CTC GAA AGG GAC T AATTTTACTG ACATGGCAAT TGAAGTGAAC GTCGGTCGTA 1464
 Arg Leu Glu Arg Asp
 445
 10 AGGTTACCGT CACG 1478

INFORMATION FOR SEQ ID NO:4:

SEQUENCE CHARACTERISTICS:

LENGTH: 445

TYPE: amino acid

TOPOLOGY: linear

MOLECULAR TYPE: protein

SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Thr Ser Ala Ser Ala Pro Ser Phe Asn Pro Gly Lys Gly Pro Gly
 1 5 10 15
 20 Ser Ala Val Gly Ile Ala Leu Leu Gly Phe Gly Thr Val Gly Thr Glu
 20 25 30
 Val Met Arg Leu Met Thr Glu Tyr Gly Asp Glu Leu Ala His Arg Ile
 35 40 45
 Gly Gly Pro Leu Glu Val Arg Gly Ile Ala Val Ser Asp Ile Ser Lys
 50 55 60
 25 Pro Arg Glu Gly Val Ala Pro Glu Leu Leu Thr Glu Asp Ala Phe Ala
 65 70 75 80
 Leu Ile Glu Arg Glu Asp Val Asp Ile Val Val Glu Val Ile Gly Gly
 85 90 95
 Ile Glu Tyr Pro Arg Glu Val Val Leu Ala Ala Leu Lys Ala Gly Lys
 100 105 110
 30 Ser Val Val Thr Ala Asn Lys Ala Leu Val Ala Ala His Ser Ala Glu
 115 120 125
 Leu Ala Asp Ala Ala Glu Ala Ala Asn Val Asp Leu Tyr Phe Glu Ala
 130 135 140
 35 Ala Val Ala Ala Ala Ile Pro Val Val Gly Pro Leu Arg Arg Ser Leu
 145 150 155 160
 Ala Gly Asp Gln Ile Gln Ser Val Met Gly Ile Val Asn Gly Thr Thr
 165 170 175
 Asn Phe Ile Leu Asp Ala Met Asp Ser Thr Gly Ala Asp Tyr Ala Asp
 180 185 190
 40 Ser Leu Ala Glu Ala Thr Arg Leu Gly Tyr Ala Glu Ala Asp Pro Thr
 195 200 205
 Ala Asp Val Glu Gly His Asp Ala Ala Ser Lys Ala Ala Ile Leu Ala
 210 215 220
 Ser Ile Ala Phe His Thr Arg Val Thr Ala Asp Asp Val Tyr Cys Glu
 225 230 235 240
 45 Gly Ile Ser Asn Ile Ser Ala Ala Asp Ile Glu Ala Ala Gln Gln Ala
 245 250 255
 Gly His Thr Ile Lys Leu Leu Ala Ile Cys Glu Lys Phe Thr Asn Lys
 260 265 270
 Glu Gly Lys Ser Ala Ile Ser Ala Arg Val His Pro Thr Leu Leu Pro
 275 280 285
 50 Val Ser His Pro Leu Ala Ser Val Asn Lys Ser Phe Asn Ala Ile Phe
 290 295 300
 Val Glu Ala Glu Ala Ala Gly Arg Leu Met Phe Tyr Gly Asn Gly Ala
 305 310 315 320

EP 0 754 756 A1

Gly Gly Ala Pro Thr Ala Ser Ala Val Leu Gly Asp Val Val Gly Ala
 325 330 335
 Ala Arg Asn Lys Val His Gly Gly Arg Ala Pro Gly Glu Ser Thr Tyr
 340 345 350
 Ala Asn Leu Pro Ile Ala Asp Phe Gly Glu Thr Thr Thr Arg Tyr His
 355 360 365
 Leu Asp Met Asp Val Glu Asp Arg Val Gly Val Leu Ala Glu Leu Ala
 370 375 380
 Ser Leu Phe Ser Glu Gln Gly Ile Ser Leu Arg Thr Ile Arg Gln Glu
 385 390 395 400
 Glu Arg Asp Asp Asp Ala Arg Leu Ile Val Val Thr His Ser Ala Leu
 405 410 415
 Glu Ser Asp Leu Ser Arg Thr Val Glu Leu Leu Lys Ala Lys Pro Val
 420 425 430
 Val Lys Ala Ile Asn Ser Val Ile Arg Leu Glu Arg Asp
 435 440 445

INFORMATION FOR SEQ ID NO:5:

SEQUENCE CHARACTERISTICS:

LENGTH: 23

TYPE: nucleic acid

STRANDEDNESS: single

TOPOLOGY: linear

MOLECULAR TYPE: other..synthetic oligonucleotide

SEQUENCE DESCRIPTION: SEQ ID NO:5:

TCGCGAAGTA GCACCTGTCA CTT 23

INFORMATION FOR SEQ ID NO:6:

SEQUENCE CHARACTERISTICS:

LENGTH: 21

TYPE: nucleic acid

STRANDEDNESS: single

TOPOLOGY: linear

MOLECULAR TYPE: other..synthetic oligonucleotide

SEQUENCE DESCRIPTION: SEQ ID NO:6:

ACGGAATTCA ATCTTACGGC C 21

INFORMATION FOR SEQ ID NO:7:

SEQUENCE CHARACTERISTICS:

LENGTH: 1643

TYPE: nucleic acid

STRANDEDNESS: double

TOPOLOGY: linear

MOLECULAR TYPE: genomic DNA

ORIGINAL SOURCE:

ORGANISM: Corynebacterium glutamicum

STRAIN: ATCC 13869

SEQUENCE DESCRIPTION: SEQ ID NO:7:

TCGCGAAGTA GCACCTGTCA CTTTGTCTC AAATATTAAA TCGAATATCA ATATACGGTC 60
 TGTTTATTGG AACGCATCCC AGTGGCTGAG ACGCATCCGC TAAAGCCCCA GGAACCCTGT 120
 GCAGAAAGAA AACACTCCTC TGGCTAGGTA GACACAGTTT ATAAAGGTAG AGTTGAGCGG 180
 GTAACGTGCA GCACGTAGAT CGAAAGGTGC ACAAAGGTGG CCCTGGTCGT ACAGAAATAT 240
 GGCGGTTCCT CGCTTGAGAG TGCGGAACGC ATTAGAAACG TCGCTGAACG GATCGTTGCC 300
 ACCAAGAAGG CTGGAAATGA TGTCGTGGTT GTCTGCTCCG CAATGGGAGA CACCACGGAT 360
 GAACTTCTAG AACTTGCAGC GGCAGTGAAT CCCGTTCCGC CAGCTCGTGA AATGGATATG 420
 CTCCTGACTG CTGGTGAGCG TATTTCTAAC GCTCTCGTCG CCATGGCTAT TGAGTCCCTT 480
 GCGCAGAAAG CTCAATCTTT CACTGGCTCT CAGGCTGGTG TGCTCACCAC CGAGCGCCAC 540
 GGAAACGCAC GCATTGTTGA CGTCACACCG GGTCTGTGTC GTGAAGCACT CGATGAGGGC 600
 AAGATCTGCA TTGTTGCTGG TTTTCAGGGT GTTAATAAAG AAACCCGCGA TGTCACCACG 660

EP 0 754 756 A1

	TTGGGTCGTG	GTGGTTCTGA	CACCACTGCA	GTTGCGTTGG	CAGCTGCTTT	GAACGCTGAT	720
	GTGTGTGAGA	TTTACTCGGA	CGTTGACGGT	GTGTATACCG	CTGACCCGCG	CATCGTTCCT	780
	AATGCACAGA	AGCTGGAAAA	GCTCAGCTTC	GAAGAAATGC	TGGAACTTGC	TGCTGTTGGC	840
5	TCCAAGATTT	TGGTGCTGCG	CAGTGTGAA	TACGCTCGTG	CATTCAATGT	GCCACTTCGC	900
	GTACGCTCGT	CTTATAGTAA	TGATCCCGGC	ACTTTGATTG	CCGGCTCTAT	GGAGGATATT	960
	CCTGTGGAAG	AAGCAGTCCT	TACCGGTGTC	GCAACCGACA	AGTCCGAAGC	CAAAGTAACC	1020
	GTTCTGGGTA	TTTCCGATAA	GCCAGGCGAG	GCTGCCAAGG	TTTTCCGTGC	GTTGGCTGAT	1080
	GCAGAAATCA	ACATTGACAT	GGTTCTGCAG	AACGTCTCCT	CTGTGGAAGA	CGGCACCACC	1140
10	GACATCACGT	TCACCTGCCC	TCGCGCTGAC	GGACGCCGTG	CGATGGAGAT	CTTGAAGAAG	1200
	CTTCAGGTTT	AGGGCAACTG	GACCAATGTG	CTTTACGACG	ACCAGGTCGG	CAAAGTCTCC	1260
	CTCGTGGGTG	CTGGCATGAA	GTCTCACCCA	GGTGTACCG	CAGAGTTCAT	GGAAGCTCTG	1320
	CGCGATGTCA	ACGTGAACAT	CGAATTGATT	TCCACCTCTG	AGATCCGCAT	TTCCGTGCTG	1380
	ATCCGTGAAG	ATGATCTGGA	TGCTGCTGCA	CGTGCATTGC	ATGAGCAGTT	CCAGCTGGGC	1440
	GGCGAAGACG	AAGCCGTCGT	TTATGCAGGC	ACCGGACGCT	AAAGTTTAA	AGGAGTAGTT	1500
15	TTACAATGAC	CACCATCGCA	GTTGTTGGTG	CAACCGGCCA	GGTCGGCCAG	GTTATGCGCA	1560
	CCCTTTTGA	AGAGCGCAAT	TTCCAGCTG	ACACTGTTCT	TTCTTTGCT	TCCCCGCGTT	1620
	CCGCAGGCCG	TAAGATTGAA	TTC				1643

INFORMATION FOR SEQ ID NO:8:

SEQUENCE CHARACTERISTICS:

LENGTH: 1643

TYPE: nucleic acid

STRANDEDNESS: double

TOPOLOGY: linear

MOLECULAR TYPE: genomic DNA

ORIGINAL SOURCE:

ORGANISM: Corynebacterium glutamicum

STRAIN: ATCC13869

FEATURE:

NAME/KEY: mat peptide

LOCATION: 217..1479

IDENTIFICATION METHOD: S

SEQUENCE DESCRIPTION: SEQ ID NO:8:

	TCGCGAAGTA	GCACCTGTCA	CTTTTGTCTC	AAATATTAAA	TCGAATATCA	ATATACGGTC	60
	TGTTTATTGG	AACGCATCCC	AGTGGCTGAG	ACGCATCCGC	TAAAGCCCCA	GGAACCCTGT	120
	GCAGAAAGAA	AACACTCCTC	TGGCTAGGTA	GACACAGTTT	ATAAAGGTAG	AGTTGAGCGG	180
35	GTAAGTGTCA	GCACGTAGAT	CGAAAGGTGC	ACAAAG	GTG	GCC CTG GTC GTA CAG	234
					Met Ala Leu Val Val Gln		
					1	5	
	AAA TAT GGC GGT TCC TCG CTT GAG AGT GCG GAA CGC ATT AGA AAC GTC						282
	Lys Tyr Gly Gly Ser Ser Leu Glu Ser Ala Glu Arg Ile Arg Asn Val						
	10	15	20				
40	GCT GAA CGG ATC GTT GCC ACC AAG AAG GCT GGA AAT GAT GTC GTG GTT						330
	Ala Glu Arg Ile Val Ala Thr Lys Lys Ala Gly Asn Asp Val Val Val						
	25	30	35				
	GTC TGC TCC GCA ATG GGA GAC ACC ACG GAT GAA CTT CTA GAA CTT GCA						378
	Val Cys Ser Ala Met Gly Asp Thr Thr Asp Glu Leu Leu Glu Leu Ala						
	40	45	50				
45	GCG GCA GTG AAT CCC GTT CCG CCA GCT CGT GAA ATG GAT ATG CTC CTG						426
	Ala Ala Val Asn Pro Val Pro Pro Ala Arg Glu Met Asp Met Leu Leu						
	55	60	65	70			
	ACT GCT GGT GAG CGT ATT TCT AAC GCT CTC GTC GCC ATG GCT ATT GAG						474
	Thr Ala Gly Glu Arg Ile Ser Asn Ala Leu Val Ala Met Ala Ile Glu						
	75	80	85				
50	TCC CTT GGC GCA GAA GCT CAA TCT TTC ACT GGC TCT CAG GCT GGT GTG						522
	Ser Leu Gly Ala Glu Ala Gln Ser Phe Thr Gly Ser Gln Ala Gly Val						
	90	95	100				

EP 0 754 756 A1

	CTC	ACC	ACC	GAG	CGC	CAC	GGA	AAC	GCA	CGC	ATT	GTT	GAC	GTC	ACA	CCG	570
	Leu	Thr	Thr	Glu	Arg	His	Gly	Asn	Ala	Arg	Ile	Val	Asp	Val	Thr	Pro	
			105					110					115				
5	GGT	CGT	GTG	CGT	GAA	GCA	CTC	GAT	GAG	GGC	AAG	ATC	TGC	ATT	GTT	GCT	618
	Gly	Arg	Val	Arg	Glu	Ala	Leu	Asp	Glu	Gly	Lys	Ile	Cys	Ile	Val	Ala	
			120					125					130				
	GGT	TTT	CAG	GGT	GTT	AAT	AAA	GAA	ACC	CGC	GAT	GTC	ACC	ACG	TTG	GGT	666
	Gly	Phe	Gln	Gly	Val	Asn	Lys	Glu	Thr	Arg	Asp	Val	Thr	Thr	Leu	Gly	
			135				140					145				150	
10	CGT	GGT	GGT	TCT	GAC	ACC	ACT	GCA	GTT	GCG	TTG	GCA	GCT	GCT	TTG	AAC	714
	Arg	Gly	Gly	Ser	Asp	Thr	Thr	Ala	Val	Ala	Leu	Ala	Ala	Ala	Leu	Asn	
					155					160					165		
	GCT	GAT	GTG	TGT	GAG	ATT	TAC	TCG	GAC	GTT	GAC	GGT	GTG	TAT	ACC	GCT	762
	Ala	Asp	Val	Cys	Glu	Ile	Tyr	Ser	Asp	Val	Asp	Gly	Val	Tyr	Thr	Ala	
				170					175					180			
15	GAC	CCG	CGC	ATC	GTT	CCT	AAT	GCA	CAG	AAG	CTG	GAA	AAG	CTC	AGC	TTC	810
	Asp	Pro	Arg	Ile	Val	Pro	Asn	Ala	Gln	Lys	Leu	Glu	Lys	Leu	Ser	Phe	
				185				190					195				
	GAA	GAA	ATG	CTG	GAA	CTT	GCT	GCT	GTT	GGC	TCC	AAG	ATT	TTG	GTG	CTG	858
	Glu	Glu	Met	Leu	Glu	Leu	Ala	Ala	Val	Gly	Ser	Lys	Ile	Leu	Val	Leu	
20			200					205					210				
	CGC	AGT	GTT	GAA	TAC	GCT	CGT	GCA	TTC	AAT	GTG	CCA	CTT	CGC	GTA	CGC	906
	Arg	Ser	Val	Glu	Tyr	Ala	Arg	Ala	Phe	Asn	Val	Pro	Leu	Arg	Val	Arg	
						220					225				230		
	TCG	TCT	TAT	AGT	AAT	GAT	CCC	GGC	ACT	TTG	ATT	GCC	GGC	TCT	ATG	GAG	954
	Ser	Ser	Tyr	Ser	Asn	Asp	Pro	Gly	Thr	Leu	Ile	Ala	Gly	Ser	Met	Glu	
25					235					240					245		
	GAT	ATT	CCT	GTG	GAA	GAA	GCA	GTC	CTT	ACC	GGT	GTC	GCA	ACC	GAC	AAG	1002
	Asp	Ile	Pro	Val	Glu	Glu	Ala	Val	Leu	Thr	Gly	Val	Ala	Thr	Asp	Lys	
				250					255					260			
	TCC	GAA	GCC	AAA	GTA	ACC	GTT	CTG	GGT	ATT	TCC	GAT	AAG	CCA	GGC	GAG	1050
	Ser	Glu	Ala	Lys	Val	Thr	Val	Leu	Gly	Ile	Ser	Asp	Lys	Pro	Gly	Glu	
30			265						270					275			
	GCT	GCC	AAG	GTT	TTC	CGT	GCG	TTG	GCT	GAT	GCA	GAA	ATC	AAC	ATT	GAC	1098
	Ala	Ala	Lys	Val	Phe	Arg	Ala	Leu	Ala	Asp	Ala	Glu	Ile	Asn	Ile	Asp	
			280				285					290					
	ATG	GTT	CTG	CAG	AAC	GTC	TCC	TCT	GTG	GAA	GAC	GGC	ACC	ACC	GAC	ATC	1146
35	Met	Val	Leu	Gln	Asn	Val	Ser	Ser	Val	Glu	Asp	Gly	Thr	Thr	Asp	Ile	
					300						305				310		
	ACG	TTC	ACC	TGC	CCT	CGC	GCT	GAC	GGA	CGC	CGT	GCG	ATG	GAG	ATC	TTG	1194
	Thr	Phe	Thr	Cys	Pro	Arg	Ala	Asp	Gly	Arg	Arg	Ala	Met	Glu	Ile	Leu	
					315					320				325			
40	AAG	AAG	CTT	CAG	GTT	CAG	GGC	AAC	TGG	ACC	AAT	GTG	CTT	TAC	GAC	GAC	1242
	Lys	Lys	Leu	Gln	Val	Gln	Gly	Asn	Trp	Thr	Asn	Val	Leu	Tyr	Asp	Asp	
				330					335					340			
	CAG	GTC	GGC	AAA	GTC	TCC	CTC	GTG	GGT	GCT	GGC	ATG	AAG	TCT	CAC	CCA	1290
	Gln	Val	Gly	Lys	Val	Ser	Leu	Val	Gly	Ala	Gly	Met	Lys	Ser	His	Pro	
			345					350					355				
45	GGT	GTT	ACC	GCA	GAG	TTC	ATG	GAA	GCT	CTG	CGC	GAT	GTC	AAC	GTG	AAC	1338
	Gly	Val	Thr	Ala	Glu	Phe	Met	Glu	Ala	Leu	Arg	Asp	Val	Asn	Val	Asn	
			360					365					370				
	ATC	GAA	TTG	ATT	TCC	ACC	TCT	GAG	ATC	CGC	ATT	TCC	GTG	CTG	ATC	CGT	1386
	Ile	Glu	Leu	Ile	Ser	Thr	Ser	Glu	Ile	Arg	Ile	Ser	Val	Leu	Ile	Arg	
			375				380				385				390		
50	GAA	GAT	GAT	CTG	GAT	GCT	GCT	GCA	CGT	GCA	TTG	CAT	GAG	CAG	TTC	CAG	1434
	Glu	Asp	Asp	Leu	Asp	Ala	Ala	Ala	Arg	Ala	Leu	His	Glu	Gln	Phe	Gln	
					395					400					405		

55

EP 0 754 756 A1

CTG GGC GGC GAA GAC GAA GCC GTC GTT TAT GCA GGC ACC GGA CGC TAA 1482
 Leu Gly Gly Glu Asp Glu Ala Val Val Tyr Ala Gly Thr Gly Arg
 410 415 420 421
 5 AGTTTAAAG GAGTAGTTTT ACAATGACCA CCATCGCAGT TGTGGTGCA ACCGGCCAGG 1542
 TCGGCCAGGT TATGCGCACC CTTTGGGAAG AGCGCAATTT CCCAGCTGAC ACTGTTCGTT 1602
 TCTTTGCTTC CCCGCGTTCC GCAGGCCGTA AGATTGAATT C 1643

INFORMATION FOR SEQ ID NO:9:

SEQUENCE CHARACTERISTICS:

LENGTH: 421

TYPE: amino acid

TOPOLOGY: linear

MOLECULAR TYPE: protein

SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met	Ala	Leu	Val	Val	Gln	Lys	Tyr	Gly	Gly	Ser	Ser	Leu	Glu	Ser	Ala
1			5					10					15		
Glu	Arg	Ile	Arg	Asn	Val	Ala	Glu	Arg	Ile	Val	Ala	Thr	Lys	Lys	Ala
			20					25					30		
Gly	Asn	Asp	Val	Val	Val	Val	Cys	Ser	Ala	Met	Gly	Asp	Thr	Thr	Asp
			35				40					45			
Glu	Leu	Leu	Glu	Leu	Ala	Ala	Ala	Val	Asn	Pro	Val	Pro	Pro	Ala	Arg
			50			55					60				
Glu	Met	Asp	Met	Leu	Leu	Thr	Ala	Gly	Glu	Arg	Ile	Ser	Asn	Ala	Leu
65					70					75				80	
Val	Ala	Met	Ala	Ile	Glu	Ser	Leu	Gly	Ala	Glu	Ala	Gln	Ser	Phe	Thr
				85					90					95	
Gly	Ser	Gln	Ala	Gly	Val	Leu	Thr	Thr	Glu	Arg	His	Gly	Asn	Ala	Arg
			100					105					110		
Ile	Val	Asp	Val	Thr	Pro	Gly	Arg	Val	Arg	Glu	Ala	Leu	Asp	Glu	Gly
		115					120					125			
Lys	Ile	Cys	Ile	Val	Ala	Gly	Phe	Gln	Gly	Val	Asn	Lys	Glu	Thr	Arg
	130					135					140				
Asp	Val	Thr	Thr	Leu	Gly	Arg	Gly	Gly	Ser	Asp	Thr	Thr	Ala	Val	Ala
145					150					155				160	
Leu	Ala	Ala	Ala	Leu	Asn	Ala	Asp	Val	Cys	Glu	Ile	Tyr	Ser	Asp	Val
				165					170					175	
Asp	Gly	Val	Tyr	Thr	Ala	Asp	Pro	Arg	Ile	Val	Pro	Asn	Ala	Gln	Lys
			180					185				190			
Leu	Glu	Lys	Leu	Ser	Phe	Glu	Glu	Met	Leu	Glu	Leu	Ala	Ala	Val	Gly
		195					200					205			
Ser	Lys	Ile	Leu	Val	Leu	Arg	Ser	Val	Glu	Tyr	Ala	Arg	Ala	Phe	Asn
	210					215					220				
Val	Pro	Leu	Arg	Val	Arg	Ser	Ser	Tyr	Ser	Asn	Asp	Pro	Gly	Thr	Leu
225					230					235				240	
Ile	Ala	Gly	Ser	Met	Glu	Asp	Ile	Pro	Val	Glu	Glu	Ala	Val	Leu	Thr
				245					250					255	
Gly	Val	Ala	Thr	Asp	Lys	Ser	Glu	Ala	Lys	Val	Thr	Val	Leu	Gly	Ile
			260					265					270		
Ser	Asp	Lys	Pro	Gly	Glu	Ala	Ala	Lys	Val	Phe	Arg	Ala	Leu	Ala	Asp
	275						280					285			
Ala	Glu	Ile	Asn	Ile	Asp	Met	Val	Leu	Gln	Asn	Val	Ser	Ser	Val	Glu
	290					295					300				
Asp	Gly	Thr	Thr	Asp	Ile	Thr	Phe	Thr	Cys	Pro	Arg	Ala	Asp	Gly	Arg
305					310					315				320	
Arg	Ala	Met	Glu	Ile	Leu	Lys	Lys	Leu	Gln	Val	Gln	Gly	Asn	Trp	Thr
				325					330					335	
Asn	Val	Leu	Tyr	Asp	Asp	Gln	Val	Gly	Lys	Val	Ser	Leu	Val	Gly	Ala
			340					345					350		

EP 0 754 756 A1

Gly Met Lys Ser His Pro Gly Val Thr Ala Glu Phe Met Glu Ala Leu
 355 360 365
 Arg Asp Val Asn Val Asn Ile Glu Leu Ile Ser Thr Ser Glu Ile Arg
 370 375 380
 Ile Ser Val Leu Ile Arg Glu Asp Asp Leu Asp Ala Ala Ala Arg Ala
 385 390 395 400
 Leu His Glu Gln Phe Gln Leu Gly Gly Glu Asp Glu Ala Val Val Tyr
 405 410 415
 Ala Gly Thr Gly Arg
 420

INFORMATION FOR SEQ ID NO:10:

SEQUENCE CHARACTERISTICS:

LENGTH: 1643

TYPE: nucleic acid

STRANDEDNESS: double

TOPOLOGY: linear

MOLECULAR TYPE: genomic DNA

ORIGINAL SOURCE:

ORGANISM: *Corynebacterium glutamicum*

STRAIN: ATCC13869

FEATURE:

NAME/KEY: mat peptide

LOCATION: 964..1479

IDENTIFICATION METHOD: S

SEQUENCE DESCRIPTION: SEQ ID NO:10:

TCGCGAAGTA GCACCTGTCA CTTTGTCTC AAATATTAAA TCGAATATCA ATATACGGTC 60
 TGTATTATGG AACGCATCCC AGTGGCTGAG ACGCATCCGC TAAAGCCCCA GGAACCCTGT 120
 GCAGAAAGAA AACACTCCTC TGGCTAGGTA GACACAGTTT ATAAAGGTAG AGTTGAGCGG 180
 GTAAGTGTCA GCACGTAGAT CGAAAGGTGC ACAAAGGTGG CCCTGGTCGT ACAGAAATAT 240
 GGCAGTTCCT CGCTTGAGAG TGCAGAACGC ATTAGAAACG TCGCTGAACG GATCGTTGCC 300
 ACCAAGAAGG CTGGAAATGA TGTCGTGGTT GTCTGCTCCG CAATGGGAGA CACCACGGAT 360
 GAACTTCTAG AACTTGCAGC GGCAGTGAAT CCCGTTCCGC CAGCTCGTGA AATGGATATG 420
 CTCCTGACTG CTGGTGAGCG TATTTCTAAC GCTCTCGTCG CCATGGCTAT TGAGTCCCTT 480
 GGCAGCAGAAG CTCAATCTTT CACTGGCTCT CAGGCTGGTG TGCTCACCAC CGAGCGCCAC 540
 GGAAACGCAC GCATTGTTGA CGTCACACCG GGTGCTGTGC GTGAAGCACT CGATGAGGGC 600
 AAGATCTGCA TTGTTGCTGG TTTTCAGGGT GTTAATAAAG AAACCCGCGA TGTCACCACG 660
 TTGGGTCGTG GTGGTTCTGA CACCACTGCA GTTGCGTTGG CAGCTGCTTT GAACGCTGAT 720
 GTGTGTGAGA TTTACTCGGA CGTTGACGGT GTGTATACCG CTGACCCGCG CATCGTTCCT 780
 AATGCACAGA AGCTGGAAAA GCTCAGCTTC GAAGAAATGC TGGAAGTTGC TGCTGTTGGC 840
 TCCAAGATTT TGGTGCTGCG CAGTGTGAA TACGCTCGTG CATTCAATGT GCCACTTCGC 900
 GTACGCTCGT CTTATAGTAA TGATCCCGGC ACTTTGATTG CCGGCTCTAT GGAGGATATT 960
 CCT GTG GAA GAA GCA GTC CTT ACC GGT GTC GCA ACC GAC AAG TCC GAA 1008
 Met Glu Glu Ala Val Leu Thr Gly Val Ala Thr Asp Lys Ser Glu
 1 5 10 15
 GCC AAA GTA ACC GTT CTG GGT ATT TCC GAT AAG CCA GGC GAG GCT GCC 1056
 Ala Lys Val Thr Val Leu Gly Ile Ser Asp Lys Pro Gly Glu Ala Ala
 20 25 30
 AAG GTT TTC CGT GCG TTG GCT GAT GCA GAA ATC AAC ATT GAC ATG GTT 1104
 Lys Val Phe Arg Ala Leu Ala Asp Ala Glu Ile Asn Ile Asp Met Val
 35 40 45
 CTG CAG AAC GTC TCC TCT GTG GAA GAC GGC ACC ACC GAC ATC ACG TTC 1152
 Leu Gln Asn Val Ser Ser Val Glu Asp Gly Thr Thr Asp Ile Thr Phe
 50 55 60
 ACC TGC CCT CGC GCT GAC GGA CGC CGT GCG ATG GAG ATC TTG AAG AAG 1200
 Thr Cys Pro Arg Ala Asp Gly Arg Arg Ala Met Glu Ile Leu Lys Lys
 65 70 75

EP 0 754 756 A1

CTT CAG GTT CAG GGC AAC TGG ACC AAT GTG CTT TAC GAC GAC CAG GTC 1248
 Leu Gln Val Gln Gly Asn Trp Thr Asn Val Leu Tyr Asp Asp Gln Val
 80 85 90 95
 5 GGC AAA GTC TCC CTC GTG GGT GCT GGC ATG AAG TCT CAC CCA GGT GTT 1296
 Gly Lys Val Ser Leu Val Gly Ala Gly Met Lys Ser His Pro Gly Val
 100 105 110
 ACC GCA GAG TTC ATG GAA GCT CTG CGC GAT GTC AAC GTG AAC ATC GAA 1344
 Thr Ala Glu Phe Met Glu Ala Leu Arg Asp Val Asn Val Asn Ile Glu
 115 120 125
 10 TTG ATT TCC ACC TCT GAG ATC CGC ATT TCC GTG CTG ATC CGT GAA GAT 1392
 Leu Ile Ser Thr Ser Glu Ile Arg Ile Ser Val Leu Ile Arg Glu Asp
 130 135 140
 GAT CTG GAT GCT GCT GCA CGT GCA TTG CAT GAG CAG TTC CAG CTG GGC 1440
 Asp Leu Asp Ala Ala Ala Arg Ala Leu His Glu Gln Phe Gln Leu Gly
 145 150 155
 15 GGC GAA GAC GAA GCC GTC GTT TAT GCA GGC ACC GGA CGC TAAAGTTTAA 1490
 Gly Glu Asp Glu Ala Val Tyr Ala Gly Thr Gly Arg
 160 165 170 172
 AGGAGTAGTT TTACAATGAC CACCATCGCA GTTGTGTTGGTG CAACCGGCCA GGTCGGCCAG 1550
 GTTATGCGCA CCCTTTTGGA AGAGCGCAAT TTCCAGCTG ACGTGTTCG TTTCTTTGCT 1610
 TCCCCGCGTT CCGCAGGCCG TAAGATTGAA TTC 1643
 20

INFORMATION FOR SEQ ID NO:11:

SEQUENCE CHARACTERISTICS:

LENGTH: 172

TYPE: amino acid

TOPOLOGY: linear

MOLECULAR TYPE: protein

SEQUENCE DESCRIPTION: SEQ ID NO:11:

Met Glu Glu Ala Val Leu Thr Gly Val Ala Thr Asp Lys Ser Glu Ala
 1 5 10 15
 Lys Val Thr Val Leu Gly Ile Ser Asp Lys Pro Gly Glu Ala Ala Lys
 20 25 30
 Val Phe Arg Ala Leu Ala Asp Ala Glu Ile Asn Ile Asp Met Val Leu
 35 40 45
 Gln Asn Val Ser Ser Val Glu Asp Gly Thr Thr Asp Ile Thr Phe Thr
 50 55 60
 Cys Pro Arg Ala Asp Gly Arg Arg Ala Met Glu Ile Leu Lys Lys Leu
 65 70 75 80
 35 Gln Val Gln Gly Asn Trp Thr Asn Val Leu Tyr Asp Asp Gln Val Gly
 85 90 95
 Lys Val Ser Leu Val Gly Ala Gly Met Lys Ser His Pro Gly Val Thr
 100 105 110
 Ala Glu Phe Met Glu Ala Leu Arg Asp Val Asn Val Asn Ile Glu Leu
 115 120 125
 40 Ile Ser Thr Ser Glu Ile Arg Ile Ser Val Leu Ile Arg Glu Asp Asp
 130 135 140
 Leu Asp Ala Ala Ala Arg Ala Leu His Glu Gln Phe Gln Leu Gly Gly
 145 150 155 160
 Glu Asp Glu Ala Val Val Tyr Ala Gly Thr Gly Arg
 165 170
 45

INFORMATION FOR SEQ ID NO:12:

SEQUENCE CHARACTERISTICS:

LENGTH: 23

TYPE: nucleic acid

STRANDEDNESS: single

TOPOLOGY: linear

MOLECULAR TYPE: other..synthetic oligonucleotide

EP 0 754 756 A1

SEQUENCE DESCRIPTION: SEQ ID NO:12:
GCCAGGCGAG CGTGCCAAGG TTT 23

5 INFORMATION FOR SEQ ID NO:13:

SEQUENCE CHARACTERISTICS:

LENGTH: 23

TYPE: nucleic acid

STRANDEDNESS: single

TOPOLOGY: linear

10 MOLECULAR TYPE: other..synthetic oligonucleotide

SEQUENCE DESCRIPTION: SEQ ID NO:13:

GCCAGGCGAG GATGCCAAGG TTT 23

15 INFORMATION FOR SEQ ID NO:14:

SEQUENCE CHARACTERISTICS:

LENGTH: 23

TYPE: nucleic acid

STRANDEDNESS: single

TOPOLOGY: linear

MOLECULAR TYPE: other..synthetic oligonucleotide

SEQUENCE DESCRIPTION: SEQ ID NO:14:

20 GCCAGGCGAG TGTGCCAAGG TTT 23

INFORMATION FOR SEQ ID NO:15:

SEQUENCE CHARACTERISTICS:

LENGTH: 23

TYPE: nucleic acid

25 STRANDEDNESS: single

TOPOLOGY: linear

MOLECULAR TYPE: other..synthetic oligonucleotide

SEQUENCE DESCRIPTION: SEQ ID NO:15:

GCCAGGCGAG TTTGCCAAGG TTT 23

30 INFORMATION FOR SEQ ID NO:16:

SEQUENCE CHARACTERISTICS:

LENGTH: 23

TYPE: nucleic acid

STRANDEDNESS: single

TOPOLOGY: linear

35 MOLECULAR TYPE: other..synthetic oligonucleotide

SEQUENCE DESCRIPTION: SEQ ID NO:16:

GCCAGGCGAG CCTGCCAAGG TTT 23

40 INFORMATION FOR SEQ ID NO:17:

SEQUENCE CHARACTERISTICS:

LENGTH: 23

TYPE: nucleic acid

STRANDEDNESS: single

TOPOLOGY: linear

45 MOLECULAR TYPE: other..synthetic oligonucleotide

SEQUENCE DESCRIPTION: SEQ ID NO:17:

GCCAGGCGAG TCTGCCAAGG TTT 23

50 INFORMATION FOR SEQ ID NO:18:

SEQUENCE CHARACTERISTICS:

LENGTH: 23

TYPE: nucleic acid

STRANDEDNESS: single

TOPOLOGY: linear

55

MOLECULAR TYPE: other..synthetic oligonucleotide
SEQUENCE DESCRIPTION: SEQ ID NO:18:
GCCAGGCGAG TATGCCAAGG TTT 23

INFORMATION FOR SEQ ID NO:19:
SEQUENCE CHARACTERISTICS:
LENGTH: 23
TYPE: nucleic acid
STRANDEDNESS: single
TOPOLOGY: linear

MOLECULAR TYPE: other..synthetic oligonucleotide
SEQUENCE DESCRIPTION: SEQ ID NO:19:
GCCAGGCGAG GTTGCCAAGG TTT 23

Claims

1. A DNA fragment which codes for homoserine dehydrogenase originating from a coryneform bacterium in which at least one of a 23rd leucine residue and a 104th valine residue as counted from its N-terminus is changed to another amino acid residue.
2. A coryneform bacterium which harbors a gene coding for mutant type homoserine dehydrogenase originating from a coryneform bacterium in which at least one of a 23rd leucine residue and a 104th valine residue as counted from its N-terminus is changed to another amino acid residue.
3. The coryneform bacterium according to claim 2, which is transformed by integrating said gene coding for mutant type homoserine dehydrogenase into chromosomal DNA by way of homologous recombination with a homoserine dehydrogenase gene on a chromosome of the coryneform bacterium.
4. The coryneform bacterium according to claim 2 or 3, wherein said other amino acid residue is a phenylalanine residue for the 23rd leucine residue, and an isoleucine residue for the 104th valine residue.
5. A coryneform bacterium wherein its homoserine dehydrogenase gene is destroyed by integrating a DNA fragment coding for a part of homoserine dehydrogenase originating from a coryneform bacterium into chromosomal DNA by way of homologous recombination with a homoserine dehydrogenase gene on a chromosome of the coryneform bacterium.
6. A coryneform bacterium which harbors in its cells recombinant DNA constructed by ligating an aspartokinase gene originating from a coryneform bacterium with a vector autonomously replicable in cells of coryneform bacteria, and expresses no wild type homoserine dehydrogenase.
7. The coryneform bacterium according to claim 6, wherein said aspartokinase gene is a gene coding for aspartokinase in which feedback inhibition by L-lysine and L-threonine is substantially desensitized.
8. A coryneform bacterium which is transformed by integrating, into chromosomal DNA of the coryneform bacterium, a gene coding for aspartokinase originating from a coryneform bacterium with desensitized feedback inhibition by L-lysine and L-threonine, and expresses no wild type homoserine dehydrogenase.
9. The coryneform bacterium according to claim 7 or 8, wherein the aspartokinase with desensitized feedback inhibition by L-lysine and L-threonine is mutant type aspartokinase in which a 279th alanine residue from the N-terminal is changed to an amino acid residue other than alanine and other than acidic amino acid in its α -subunit, and a 30th alanine residue is changed to an amino acid residue other than alanine and other than acidic amino acid in its β -subunit.
10. The coryneform bacterium according to any one of claims 6-9, which is transformed by integrating, into its chromosome, a mutant type homoserine dehydrogenase gene for homoserine dehydrogenase originating from a coryneform bacterium in which at least one of a 23rd leucine residue and a 104th valine residue as counted from the N-

terminal is changed to another amino acid residue, by way of homologous recombination with a homoserine dehydrogenase gene on the chromosome of the coryneform bacterium, and thus expresses no wild type homoserine dehydrogenase.

- 5 11. The coryneform bacterium according to any one of claims 6-9, which has its homoserine dehydrogenase gene destroyed by integrating, into its chromosome, a DNA fragment coding for a part of homoserine dehydrogenase originating from a coryneform bacterium, by way of homologous recombination with a homoserine dehydrogenase gene on the chromosome of the coryneform bacterium, and expresses no wild type homoserine dehydrogenase.
- 10 12. A method of producing L-lysine comprising the steps of cultivating the coryneform bacterium according to any one of claims 2-11 in an appropriate medium, producing and accumulating L-lysine in a culture thereof, and collecting L-lysine from the culture.

15

20

25

30

35

40

45

50

55

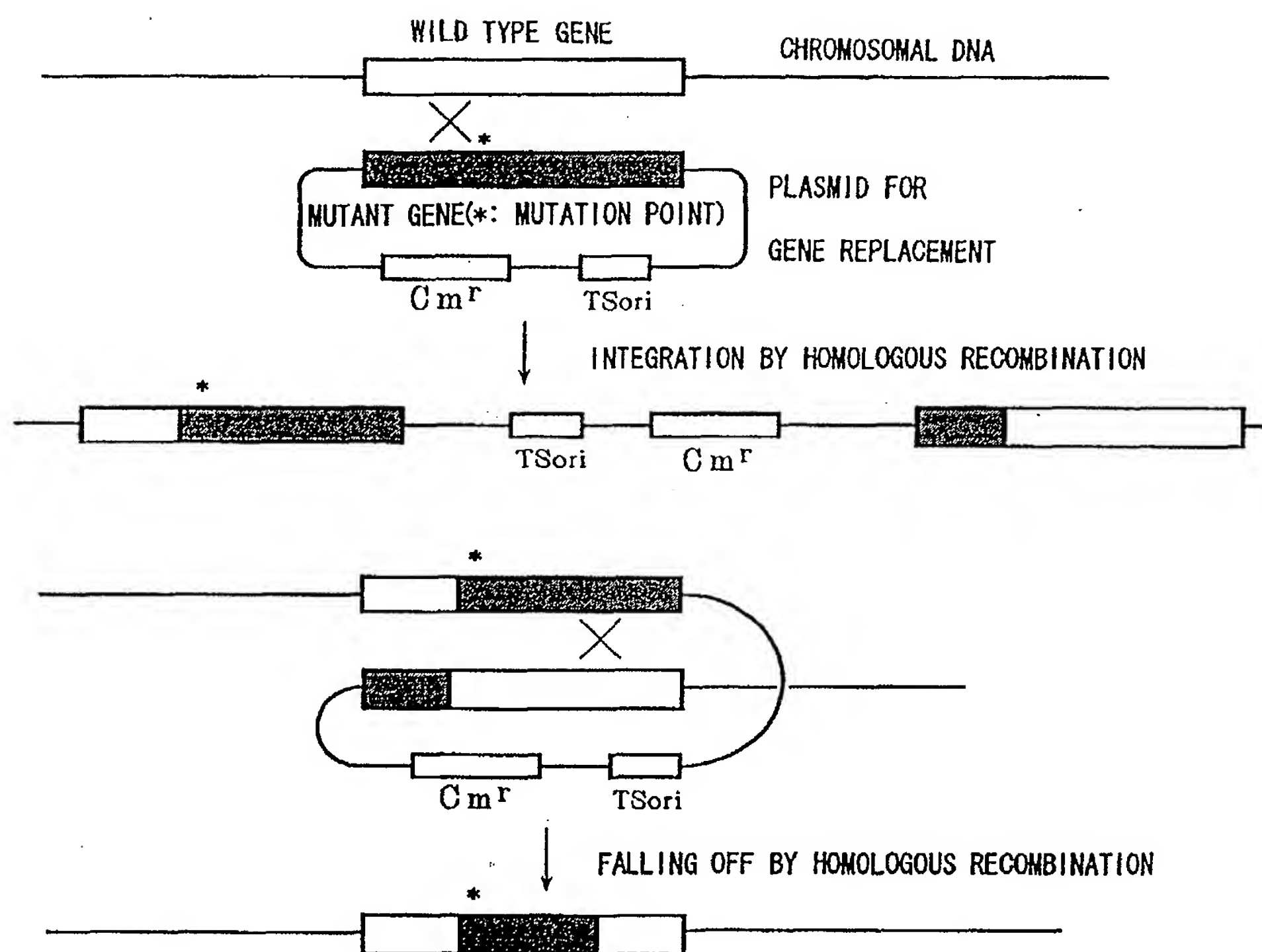


Fig. 1

EP 0 754 756 A1

BL	M	T	S	A	S	A	P	S	*	F	N	P	G	K	G	P	G	S	A	V	G	I	A	L	L	G	F	G	T	V	G	T	E	32
BS													M	K	-	-	-	-	A	I	R	V	G	L	L	G	L	G	T	V	G	S	G	17
E1	V	R	V	T	H	Q	M	L	F	N	T	D	Q	-	-	-	-	V	I	E	V	F	V	I	G	V	G	G	V	G	G	A	479	
E2	I	Q	G	L	H	Q	S	V	F	R	A	E	K	-	-	-	-	R	I	G	L	V	L	F	G	K	G	N	I	G	S	R	471	
BL	V	M	R	L	M	T	E	Y	G	D	E	L	A	H	R	I	G	G	P	L	E	V	R	G	I	A	-	V	S	D	I	S	63	
BS	V	V	K	I	I	Q	D	H	Q	D	K	L	M	H	Q	V	G	C	P	-	-	V	-	T	I	K	K	V	-	-	-	-	42	
E1	L	L	E	Q	L	K	R	Q	Q	S	W	L	K	N	K	-	-	H	I	D	L	R	V	C	-	G	V	A	N	-	S	506		
E2	W	L	E	L	F	A	R	E	Q	S	T	L	S	A	R	-	-	T	G	F	E	F	V	L	A	G	V	V	D	-	S	499		
BL	K	-	-	-	-	P	R	E	G	V	A	P	E	L	L	T	E	D	A	F	A	L	I	E	R	E	D	V	D	I	V	90		
BS	-	-	-	-	-	-	-	-	-	-	-	L	V	K	D	L	E	K	K	R	E	V	D	L	P	K	E	V	L	T	T	62		
E1	K	-	A	L	L	T	E	V	H	G	L	N	L	E	N	W	Q	E	E	L	A	Q	A	K	E	P	P	N	L	G	R	L	537	
E2	R	R	S	L	L	S	Y	-	D	G	L	D	A	S	R	A	L	A	F	F	N	D	E	A	V	E	Q	D	E	E	S	L	530	
BL	V	E	V	I	G	G	I	E	Y	P	R	E	-	V	V	L	A	A	-	-	-	-	-	-	-	-	-	-	-	-	-	L	108	
BS	E	-	V	Y	D	V	I	D	D	P	D	V	D	V	V	I	E	V	I	G	G	V	E	Q	T	K	Q	Y	L	V	D	A	93	
E1	I	R	L	V	K	E	Y	H	L	L	N	-	P	V	I	V	N	C	T	S	S	Q	A	V	A	D	Q	Y	A	D	P	L	568	
E2	F	-	L	W	M	R	A	H	P	Y	D	D	L	V	V	L	D	V	T	A	S	Q	Q	L	A	D	Q	Y	L	D	F	A	561	
BL	-	K	A	G	K	S	V	V	T	A	N	K	-	A	L	V	A	A	H	S	A	E	-	-	L	A	D	A	A	E	A	A	136	
BS	L	R	S	K	K	H	V	V	T	A	N	K	D	L	M	A	-	-	-	-	V	Y	G	S	E	L	L	A	E	A	K	E	121	
E1	-	R	E	G	F	H	V	V	T	P	N	K	K	A	N	T	S	S	M	D	Y	Y	H	Q	L	R	Y	A	A	E	K	S	599	
E2	-	S	H	G	F	H	V	I	S	A	N	K	L	A	G	A	S	D	S	N	K	Y	R	Q	I	H	D	A	F	E	K	T	592	
BL	N	V	D	-	L	Y	F	E	A	A	V	A	A	A	I	P	V	V	G	P	L	R	R	S	L	-	A	G	D	Q	I	Q	166	
BS	N	G	C	D	I	Y	F	E	A	S	V	A	G	G	I	P	I	L	R	T	L	E	E	G	L	S	S	-	D	R	I	T	152	
E1	R	R	K	F	L	Y	-	D	I	N	V	G	A	G	L	P	V	I	E	N	L	Q	N	L	L	N	A	G	D	E	L	M	630	
E2	G	R	H	W	L	Y	-	N	A	T	V	G	A	G	L	P	I	N	H	T	V	R	D	L	I	D	S	G	D	T	I	L	623	
BL	S	V	M	G	I	V	N	G	T	T	N	F	I	L	D	A	M	-	D	S	T	G	A	D	Y	A	D	S	L	A	E	A	197	
BS	K	M	M	G	I	V	N	G	T	T	N	F	I	L	T	K	M	I	K	E	K	S	P	-	Y	E	-	-	-	-	E	V	179	
E1	K	F	S	G	I	L	S	G	S	L	S	Y	I	F	G	K	-	L	D	E	G	M	S	-	F	S	-	-	-	-	E	A	656	
E2	S	I	S	G	I	F	S	G	T	L	S	W	L	F	L	Q	-	F	D	G	S	V	P	-	F	T	-	-	-	-	E	L	649	
BL	T	R	-	-	-	L	G	Y	A	E	A	D	P	T	A	D	V	E	G	H	D	A	A	S	K	A	A	I	L	A	S	225		
BS	L	K	E	A	Q	D	L	G	F	A	E	A	D	P	T	S	D	V	E	G	L	D	A	A	R	K	M	A	I	L	A	R	211	
E1	T	R	L	A	R	E	M	G	Y	T	E	P	D	P	R	D	D	L	S	G	M	D	V	A	R	K	L	L	I	L	A	R	688	
E2	V	D	Q	A	W	Q	Q	G	L	T	E	P	D	P	R	D	D	L	S	G	K	D	V	S	R	K	L	V	I	L	A	R	681	

Fig. 2

BL: *Brevibacterium lactofermentum* BS: *Bacillus subtilis*

E1: *E. coli* HDI

E2: *E. coli* HDII

*: amino acid common to three species

** : amino acid common to four species

EP 0 754 756 A1

BL	I	A	F	H	T	R	V	T	A	D	D	V	Y	C	E	G	I	S	N	I	S	A	A	D	I	E	-	A	A	Q	Q	A	256
BS	L	-	G	F	S	M	N	V	D	L	E	D	V	K	V	K	G	I	S	Q	I	T	D	E	D	I	S	F	-	-	-	-	238
E1	E	T	G	R	E	L	E	L	A	D	I	E	I	E	P	V	L	P	A	E	F	N	A	E	G	D	V	A	A	F	M	A	720
E2	E	A	G	Y	N	I	E	P	D	Q	V	R	V	E	S	L	V	P	A	-	H	C	E	G	G	S	I	D	H	F	F	E	712
BL	G	H	T	I	-	-	-	-	-	-	-	-	-	-	-	-	K	L	L	A	I	C	E	K	F	T	N	K	E	G	K	S	276
BS	-	-	-	-	-	-	-	-	-	-	-	S	K	R	L	C	Y	T	M	K	L	I	G	I	A	Q	R	D	G	S	K	258	
E1	N	L	S	Q	L	D	D	L	F	A	A	R	V	A	K	A	R	D	E	G	K	V	L	R	Y	V	G	N	I	D	E	D	752
E2	N	G	D	E	L	N	E	Q	M	V	Q	R	L	E	A	A	R	E	M	G	L	V	L	R	Y	V	A	R	F	D	A	N	744
BL	A	I	S	A	R	V	H	P	T	L	L	P	V	S	H	P	L	A	S	V	N	K	S	F	N	A	I	F	V	E	A	E	308
BS	I	E	-	V	S	V	Q	P	T	L	L	P	D	H	H	P	L	S	A	V	H	N	E	F	N	A	V	Y	V	Y	G	E	289
E1	G	V	-	C	R	V	K	I	A	E	V	D	G	N	D	P	L	F	K	V	K	N	G	E	N	A	L	A	F	Y	S	H	783
E2	G	K	-	A	R	V	G	V	E	A	V	R	E	D	H	P	L	A	S	L	L	P	C	D	N	V	F	A	I	E	S	R	775
BL	A	A	G	R	L	M	F	Y	-	-	G	N	G	A	G	G	A	P	T	A	S	A	V	L	G	D	V	V	G	A	A	R	338
BS	A	V	G	E	T	M	F	Y	-	-	G	P	G	A	G	S	M	P	T	A	T	S	V	V	S	D	L	V	A	V	M	K	319
E1	Y	Y	Q	P	L	P	L	V	L	R	G	Y	G	A	G	N	D	V	T	A	A	G	V	P	A	D	L	L	R	T	L	S	814
E2	W	Y	R	D	N	P	L	V	I	R	G	P	G	A	G	R	D	V	T	A	G	A	I	Q	S	D	I	N	R	L	A	Q	806
BL	N	K	V	H	G	R	A	P	G	E	S	T	Y	A	N	L	P	I	A	D	F	G	E	T	T	T	R	Y	H	L	D	370	
BS	N	M	R	L	G	V	T	G	N	S	F	V	G	P	Q	Y	E	K	N	M	K	S	P	S	D	I	Y	A	Q	Q	F	L	351
E1	W	K	-	L	G	V																										817	
E2	L	-	-	L																												806	
BL	M	D	-	V	E	D	R	V	G	V	L	A	E	L	A	S	L	F	S	E	Q	G	I	S	L	R	T	I	R	Q	E	E	401
BS	R	I	H	V	K	D	E	V	G	S	F	S	K	I	T	S	V	F	S	E	R	G	V	S	F	E	K	I	L	Q	L	P	383
BL	R	D	D	-	-	-	D	A	R	L	I	V	V	T	H	S	A	L	E	S	D	L	S	R	T	V	E	L	L	K	A	K	430
BS	I	K	G	H	D	E	L	A	E	I	V	I	V	T	H	H	T	S	E	A	D	F	S	D	I	L	Q	N	L	N	D	L	415
BL	P	V	V	K	A	I	N	S	V	I	R	L	E	R	D																		445
BS	E	V	V	Q	E	V	K	S	T	Y	R	V	E	G	N	G	W	S															433

Fig. 3

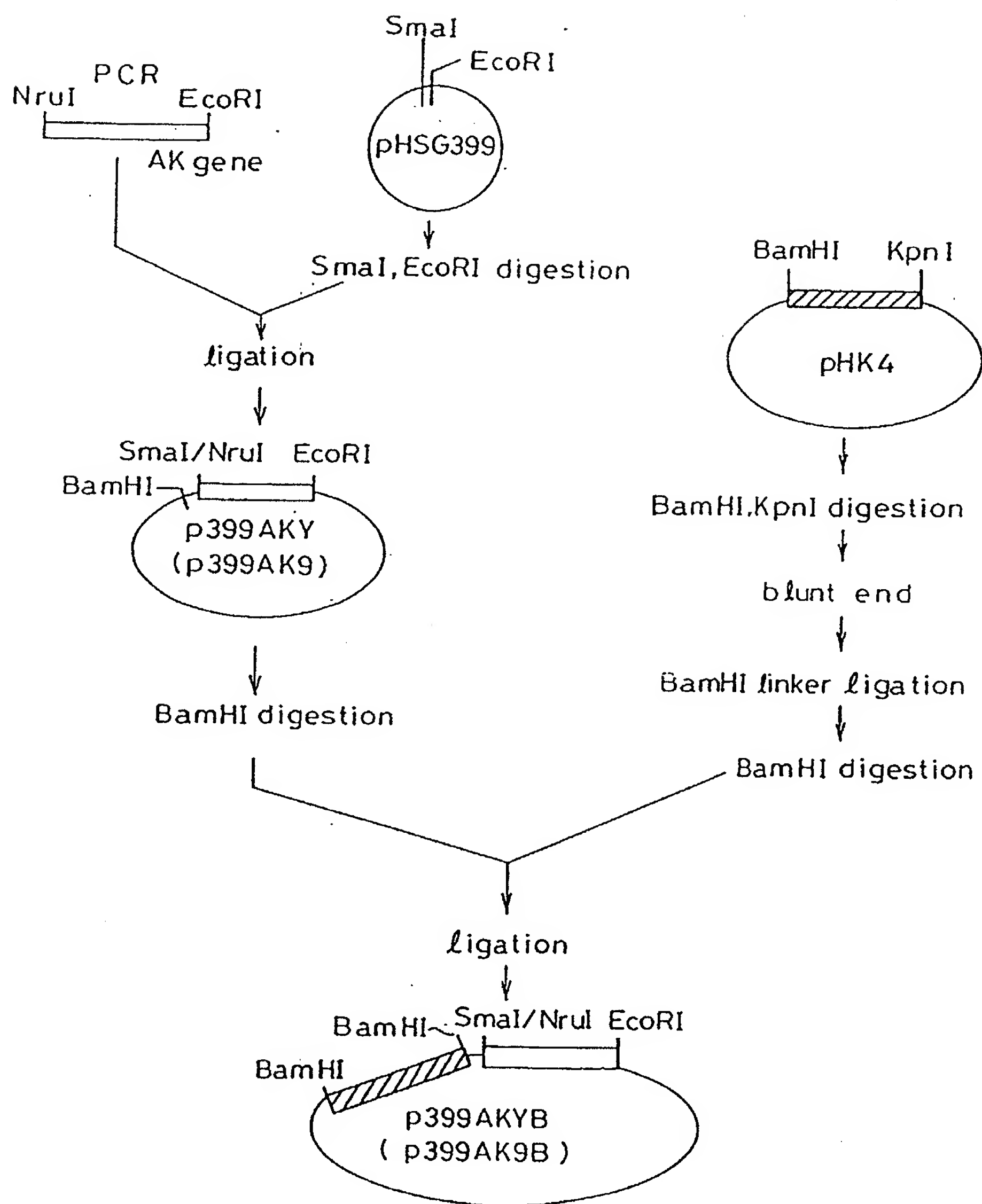


Fig. 4

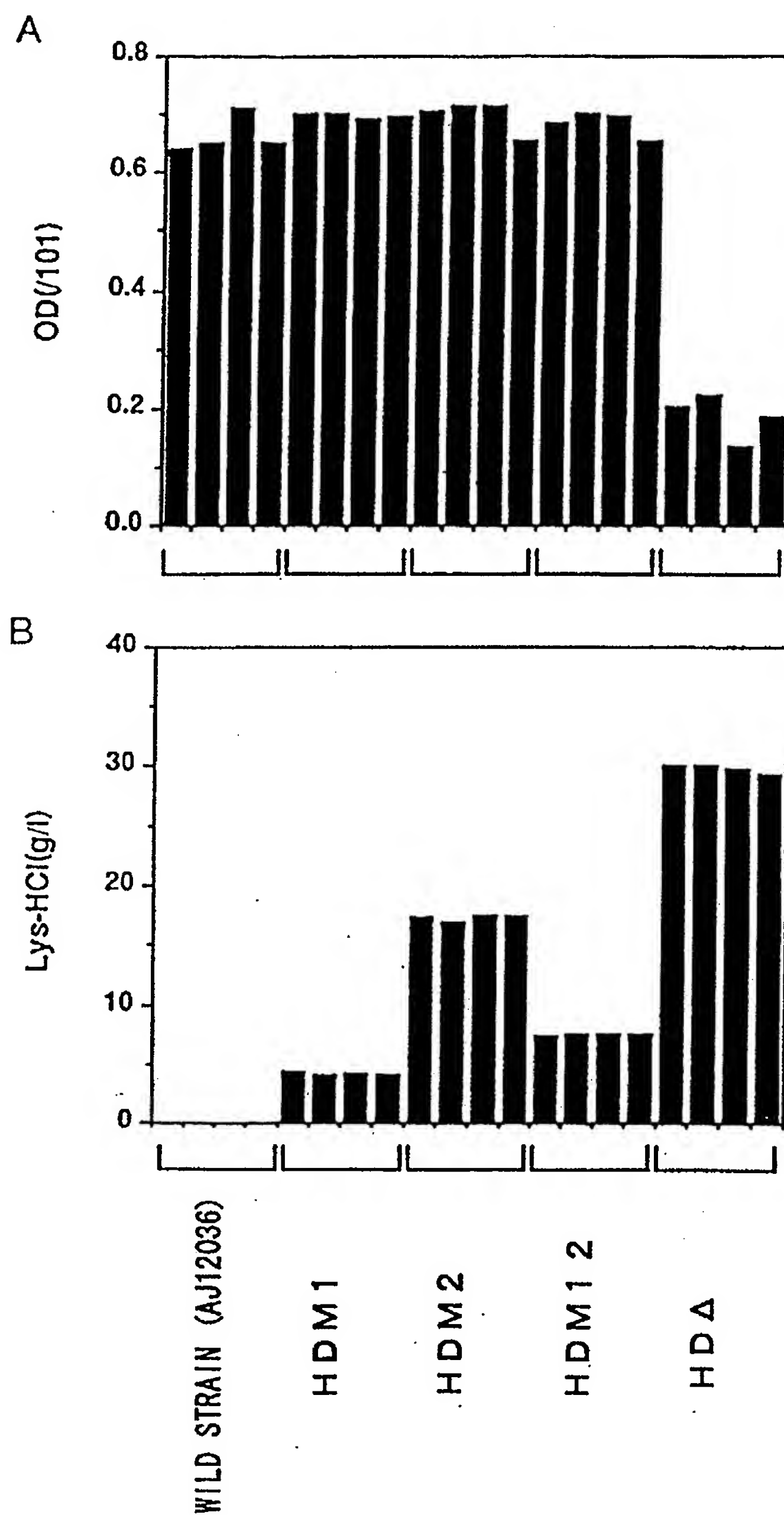


Fig. 5

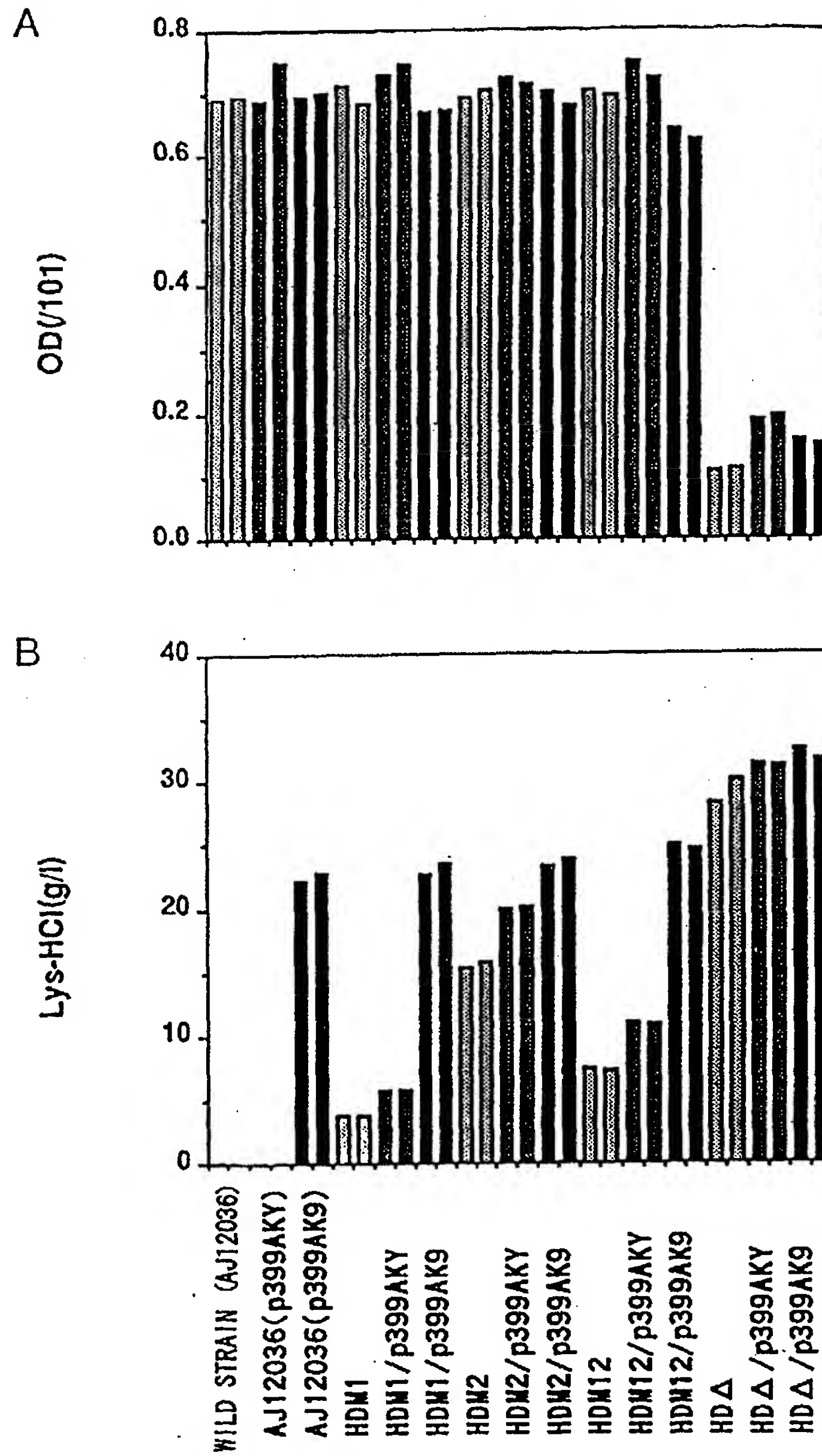


Fig. 6

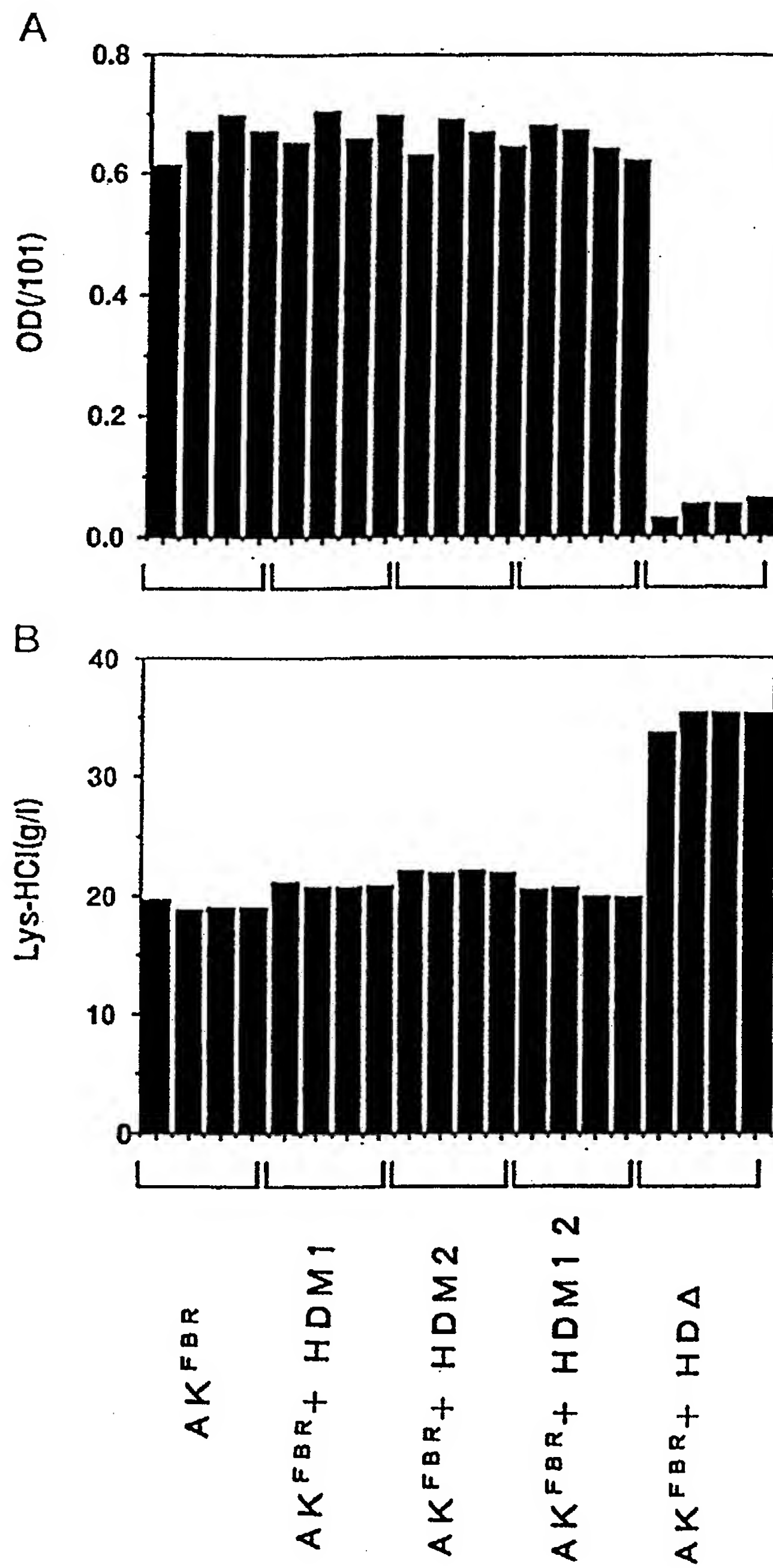


Fig. 7

INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP95/00268

A. CLASSIFICATION OF SUBJECT MATTER		
Int. C1 ⁶ C12N15/53, C12P13/08		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED		
Minimum documentation searched (classification system followed by classification symbols)		
Int. C1 ⁶ C12N15/53, C12P13/08		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)		
CAS BIOSIS WPI, WPI/L		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y/A	Mol. Microbiol. Vol. 2, No. 1 (1988), Peoples, O. P. et al. "Nucleotide sequence and fine structural analysis of the Corynebacterium glutamicum hom-thr B operon" p. 63-72	5, 6, 7, 8/ 1-4, 9-12
Y/A	Mol. Microbiol. Vol. 5, No. 5 (1991), Kalinowski, J. et al. "Genetic biochemical analysis of the aspartokinase from Corynebacterium glutamicum" p. 1197-1204	6, 7, 8/9-12
Y/A	JP, A, 3-219885 (Degussa AG.), September 27, 1991 (27. 09. 91), & EP, A1, 387527	6, 7, 8/12
A	J, Gen. Appl. Microbiol. Vol. 7, No. 3 (1961) Nakayama et al. p. 145-154	1 - 12
Y/A	EP, A, 435132 (FORSCHUNGSZENT JUELICH GMBH), July 3, 1991 (03. 07. 91), & DE, A, 3943117	6, 7, 8/9-12
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family		
Date of the actual completion of the international search		Date of mailing of the international search report
April 13, 1995 (13. 04. 95)		May 2, 1995 (02. 05. 95)
Name and mailing address of the ISA/ Japanese Patent Office		Authorized officer
Facsimile No.		Telephone No.

Form PCT/ISA/210 (second sheet) (July 1992)